

# INTEGRATED DETECTION and MANIPULATION of PARAMAGNETIC MICROSPHERES

Mark Tondra, NVE Corp. Eden Prairie, MN  
EMD Biosciences / Estapor Meeting, June 17, 2005

- Magnetic Biosensor Concept: Biochemical binding + magnetic detection
- Magnetoresistive biosensor fabrication
- Detection and Manipulation Examples: Scanning, Immobilized and Flowing Labels, Flow Sorting

# NVE Corporation

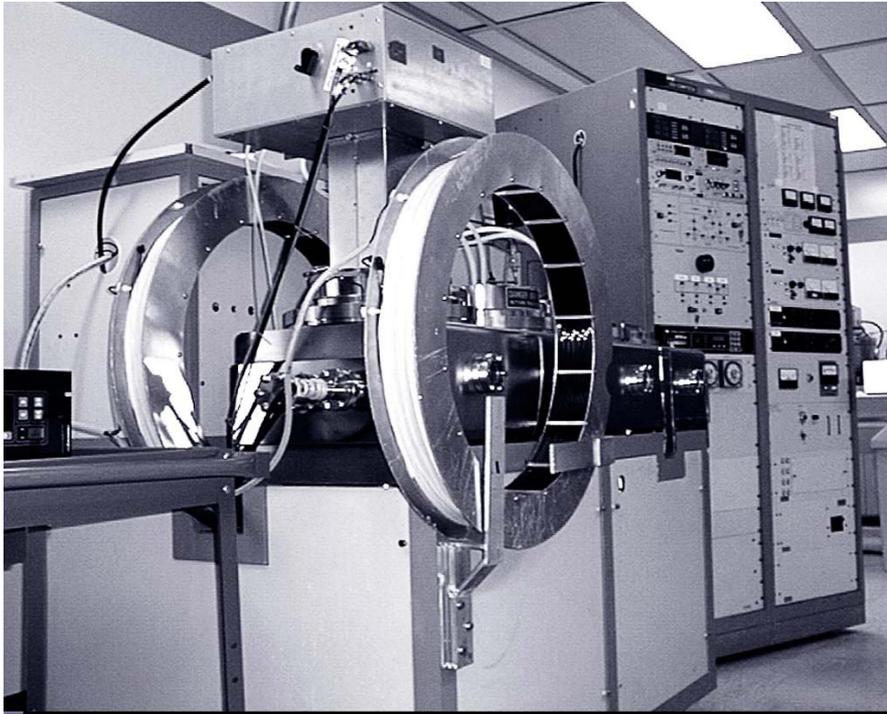
- Founded in 1989 as Nonvolatile Electronics
- Now traded on NASDAQ as NVEC
- About 60 employees
- 6000 ft<sup>2</sup> clean room (class 100)
- Specialize in integrated magnetoresistive devices
- FY '05 revenues ~\$12M
  - Magnetic Sensors [biomedical, industrial]
  - Digital signal couplers
  - Contract and Govt. Research and Development



# NVE Highlights

- Licensed by Honeywell in 1989 to produce Magnetoresistive Random Access Memory (MRAM)
- Introduced world's first Giant Magnetoresistance (GMR) product in 1996
- GMR Signal isolator product introduced in 1999
- Research & Development Thrusts
  - Biosensors
  - MRAM
  - Low-Field Magnetometers
  - Non-destructive Evaluation
  - Electronics and systems
- About 60 employees

# NVE Fabrication Facility



Comptech

PE 2400



# Acknowledgements

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IOWA STATE  
UNIVERSITY



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Cy Tamanaha  
Shaun Mulvaney

[www.nve.com/~markt/biomagnetics](http://www.nve.com/~markt/biomagnetics)



# Motivation for Current R&D Efforts

**Military / Homeland Defense** wants bioassays that are:

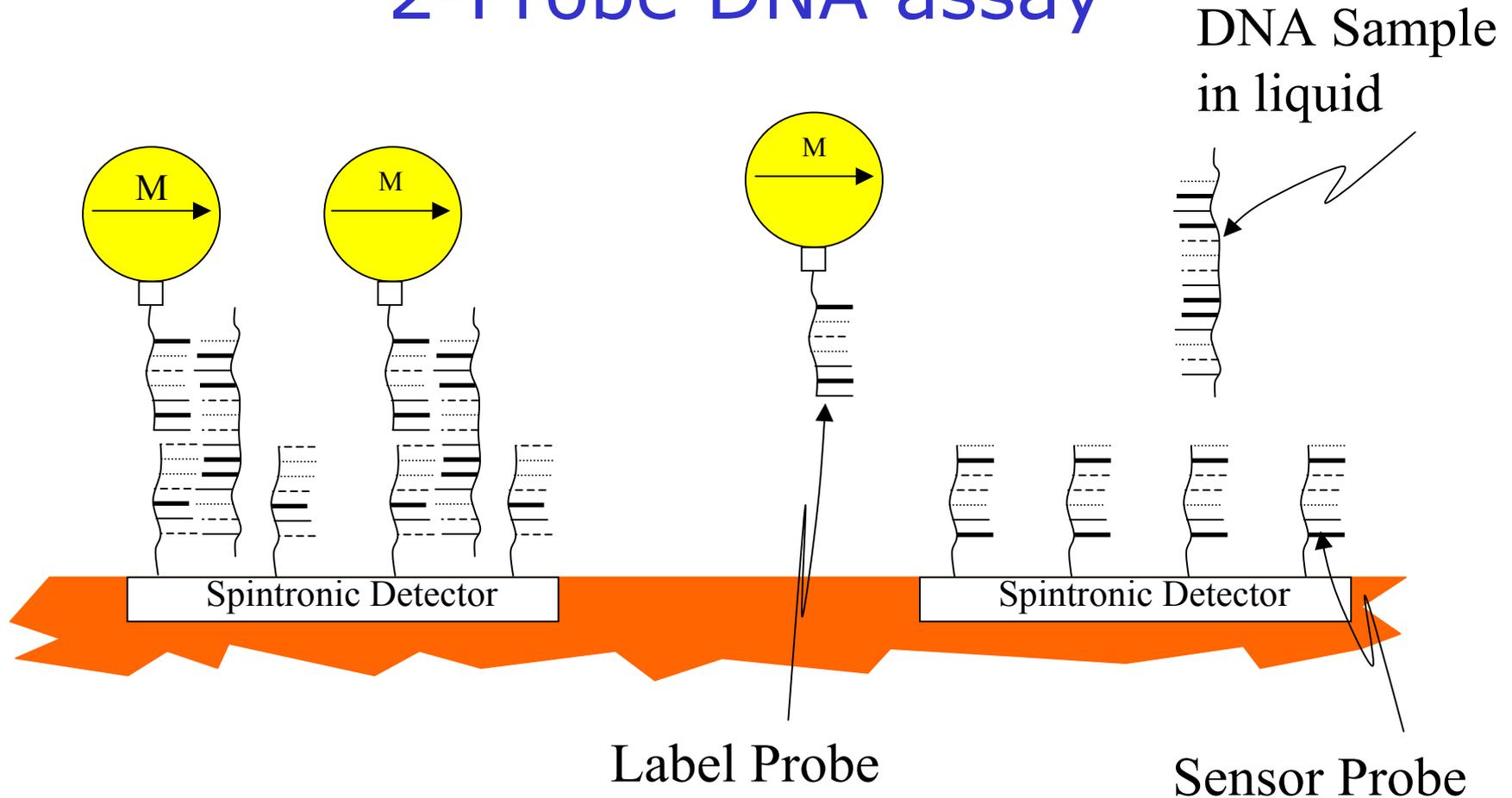
- Rugged
- Lightweight / handheld
- Cheap
- Rapid
- (Also: highly sensitive and specific, multi-functional, fool-proof, etc.)
- Readers, sensors, and fluidics must be mass-manufacturable.

# Status of Development

- Developing some great magnetic detection tools
- Working with excellent university and Lab. Partners
- Looking for commercial partners to provide access to the marketplace for specific diagnostic products
- Mass-manufacturable sensor process,  $\ll \$1 / \text{mm}^2$

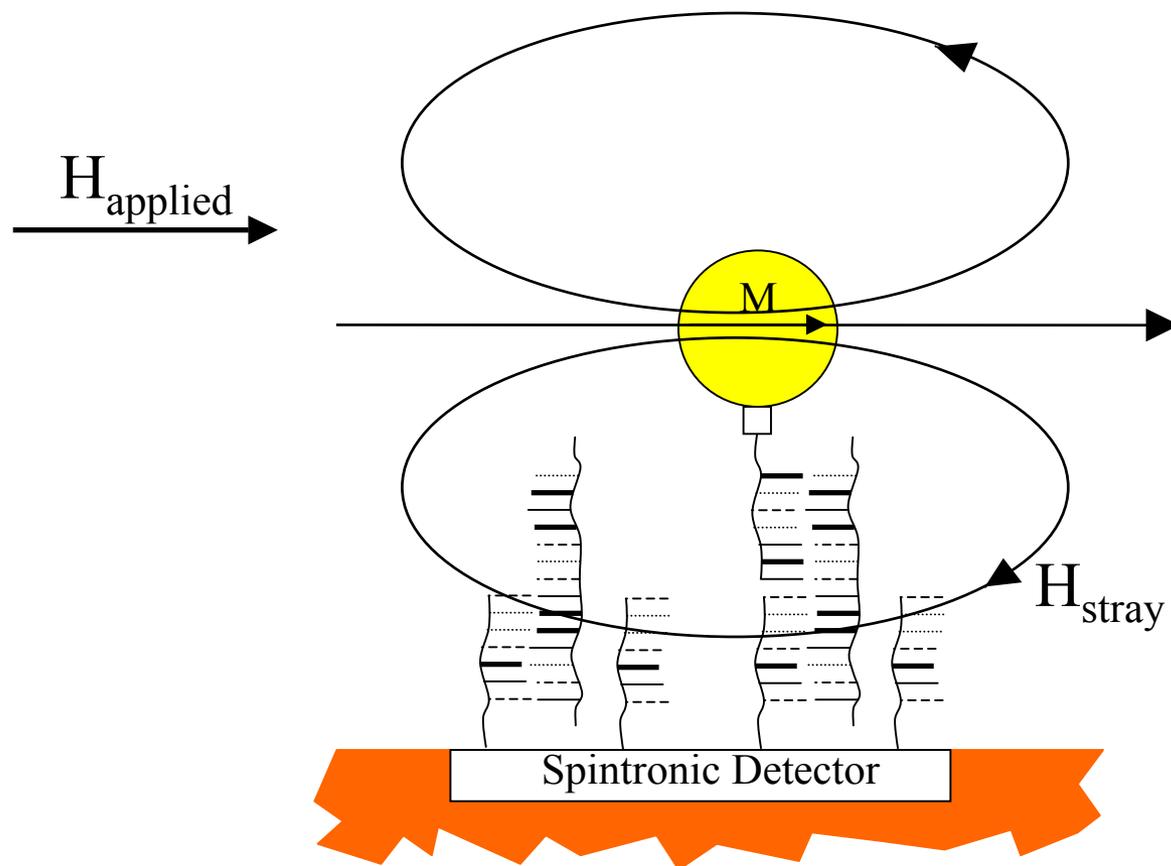
# Magnetic Nanolabel Example:

## 2-Probe DNA assay



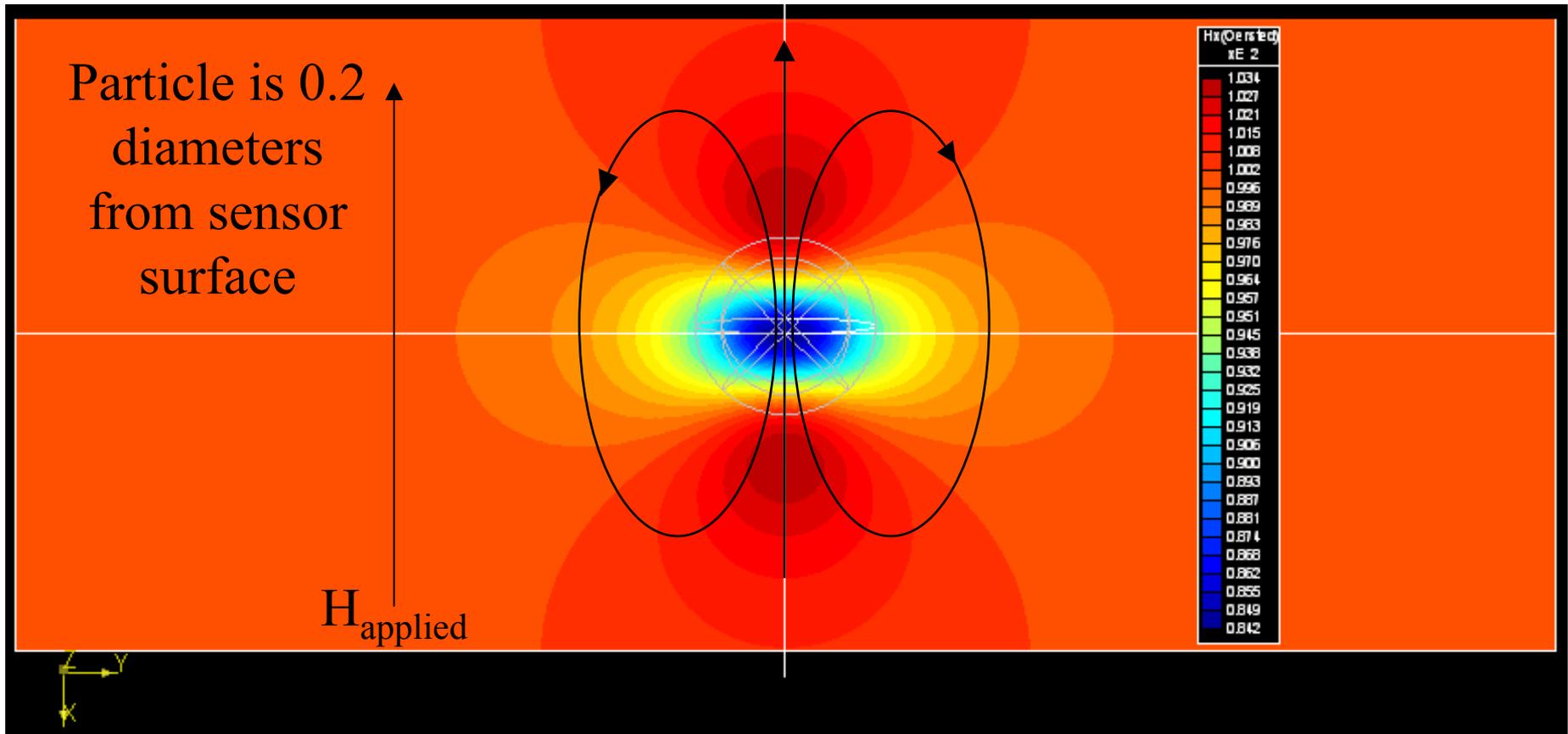
# Stray Fields from Bound Magnetic Nanolabel

Detector sees  $H_{\text{Total}} = H_{\text{applied}} + H_{\text{stray}}$  along a designed sense-axis



# Model prediction for single label – top view

Total ( $H_{\text{app}} + H_{\text{stray}}$ ) sense-axis field in sensor plane from single particle



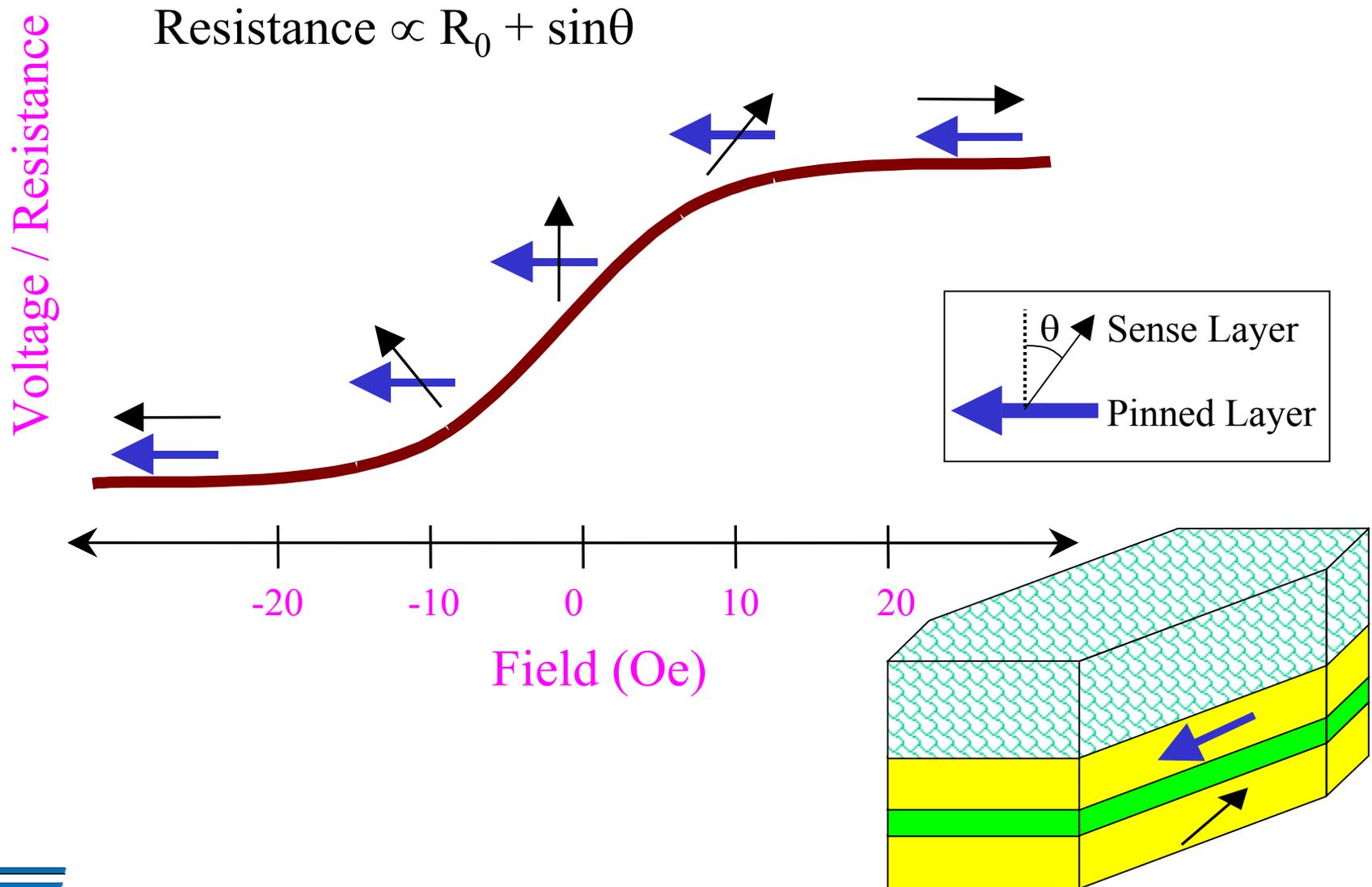
# Detector Design Parameters

- 1 – Field of view: (Sensor Area) x (*Label Diameter*)
- 2 – Dynamic Range: Max / Min # of labels detectable
- 3 – Sensitivity: microvolts / label
- 4 – Noise: (microvolts / Hz<sup>0.5</sup>) x (Bandwidth(Hz))<sup>0.5</sup>

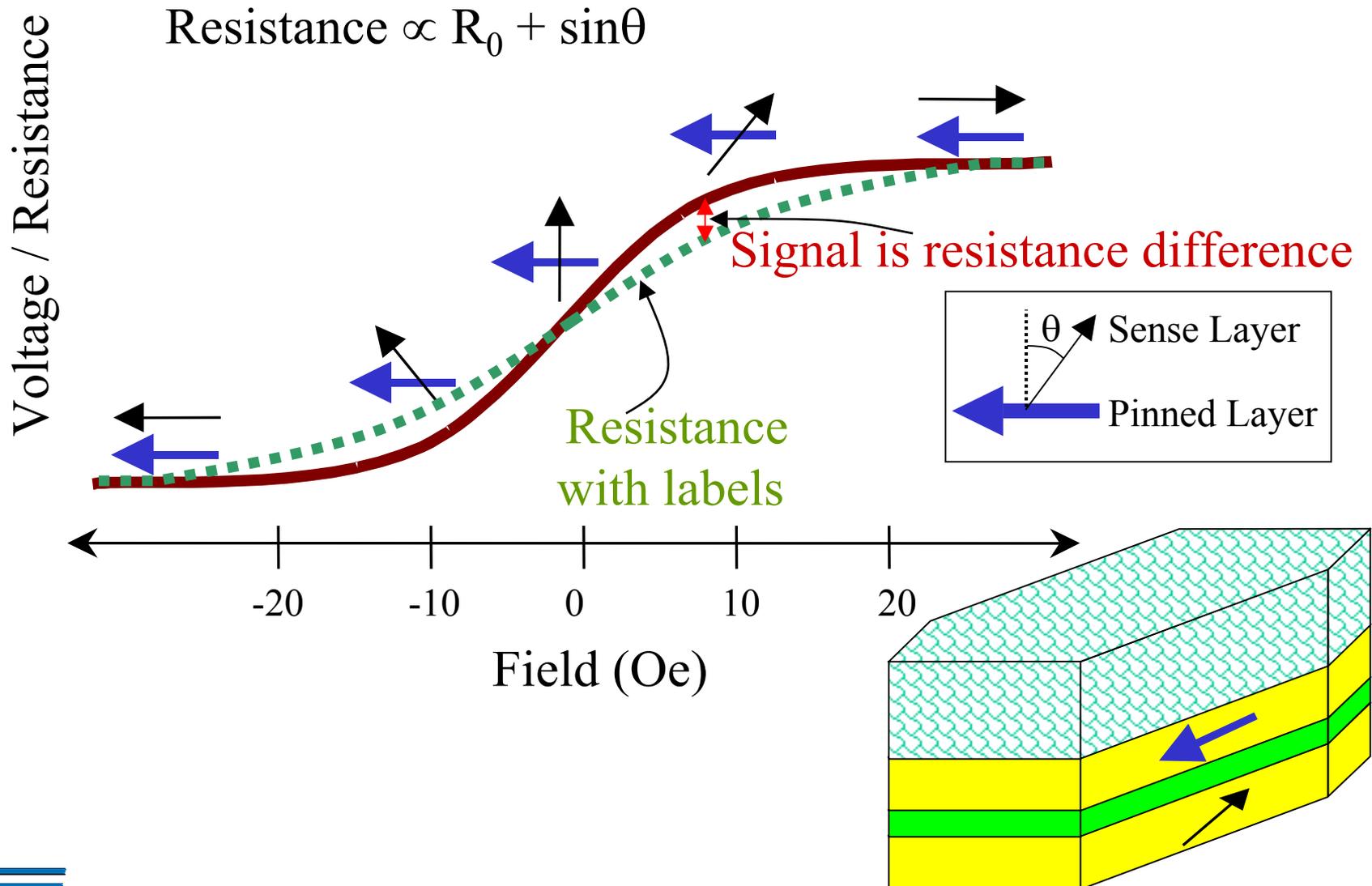
# GMR Biosensor Detection Capabilities

- 1 - Single label detection – small sensor required
- 2 – Combine many small sensors together to make 200 micron diameter sensor (match spot size)
- 3 – Field of view: (Sensor Area) x (*Label Diameter*)
- 4 – Dynamic Range of detected labels can be  $\gg 3$  logs

# Idealized Spin Valve Transfer Curve



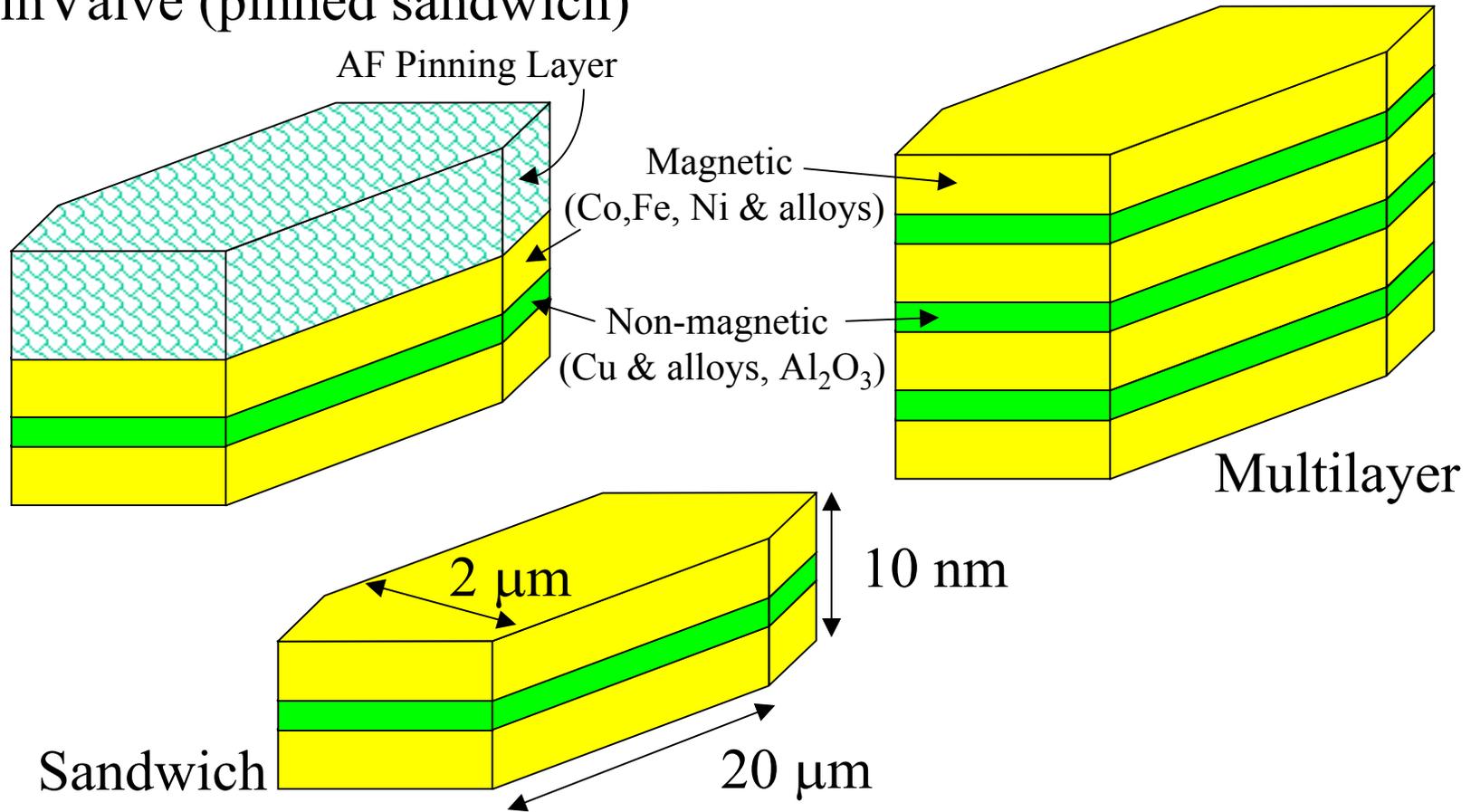
# Resistance when labels are present



# Magnetoresistive Material Structures

Resistance change due to Spin Dependent Electron Transport

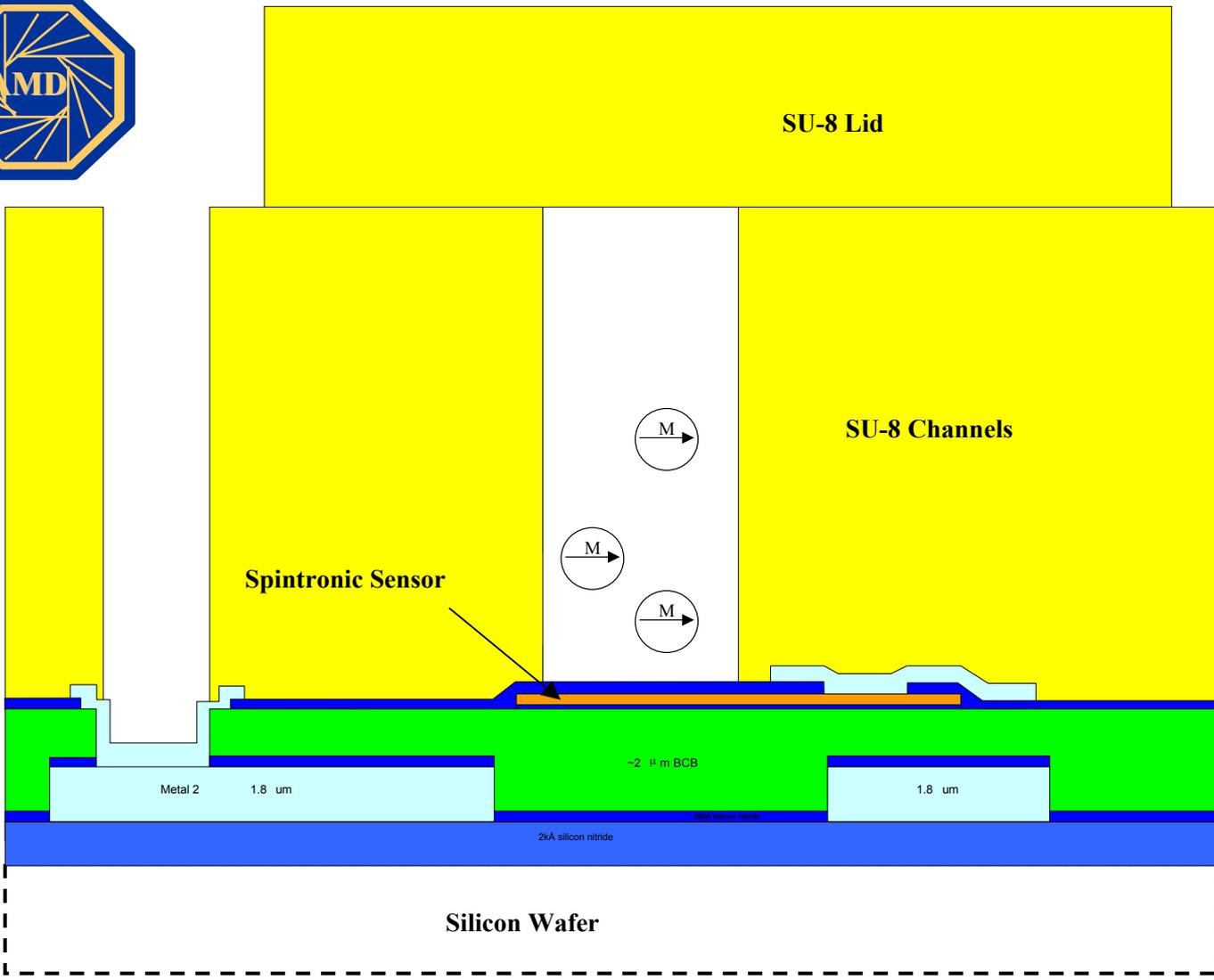
Spin Valve (pinned sandwich)



# Sensors produced in semicond. wafer fab

- Layer by layer deposition, photolithography, etch
- 1 150mm diameter wafer can deliver >10,000 sensors
- Integrated circuitry is easily added to basic sensor

# SU-8 Lid using wafer-scale bonding method



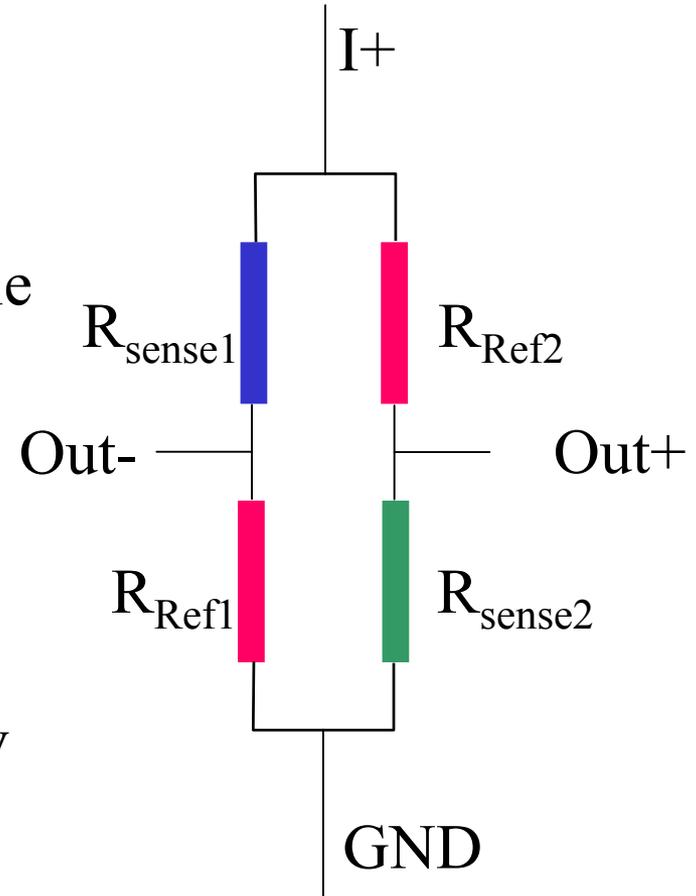
# Wheatstone Bridge Design

Reference resistors for field and temperature compensation

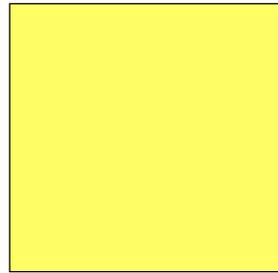
2 Sense resistors that experience the sample or field being measured

Reference resistors do not experience sample

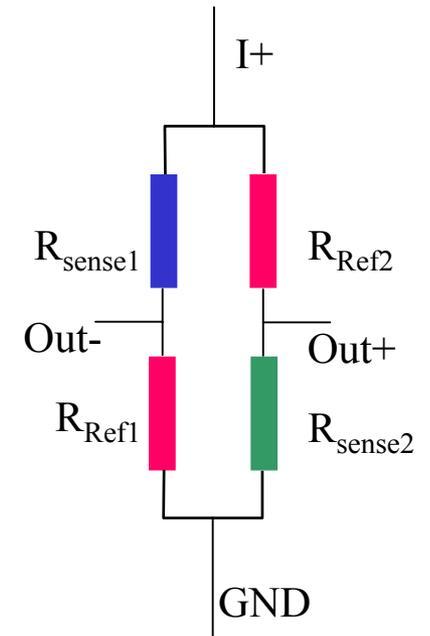
$V_{out} = Out+ - Out-$ . If there is nothing to make the sense resistors behave differently than the refs,  $V_{out} = 0$  Volts



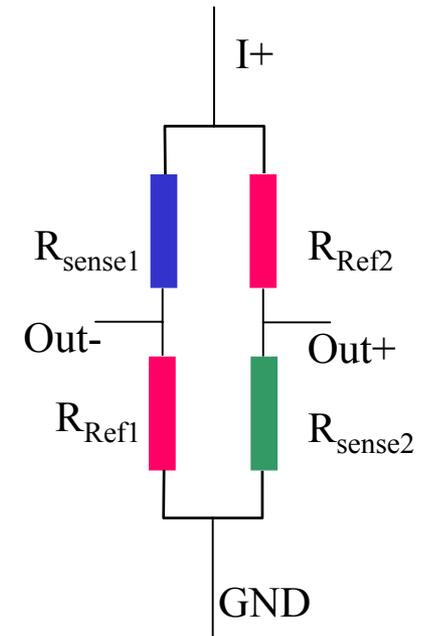
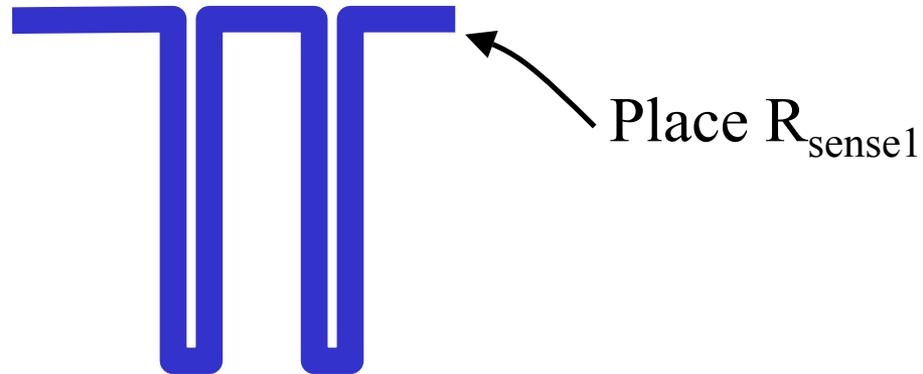
# Layout: Define Sensing Region



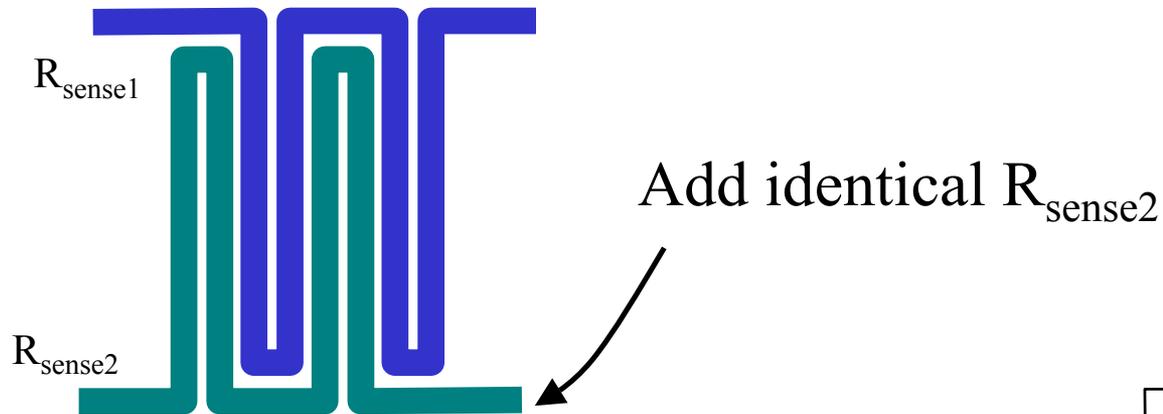
100  $\mu\text{m}$  x 100  $\mu\text{m}$  sense region



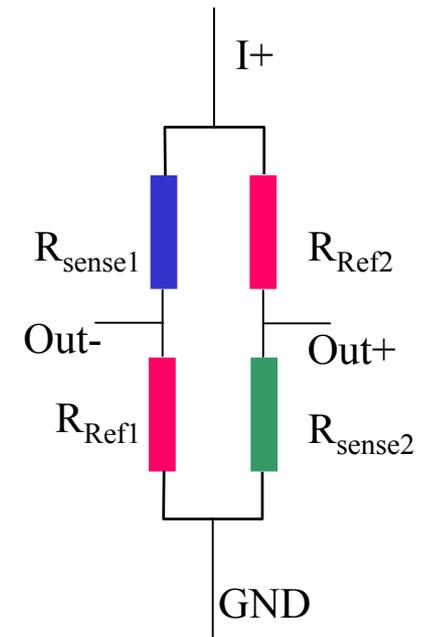
# Layout: Place Sense Resistors



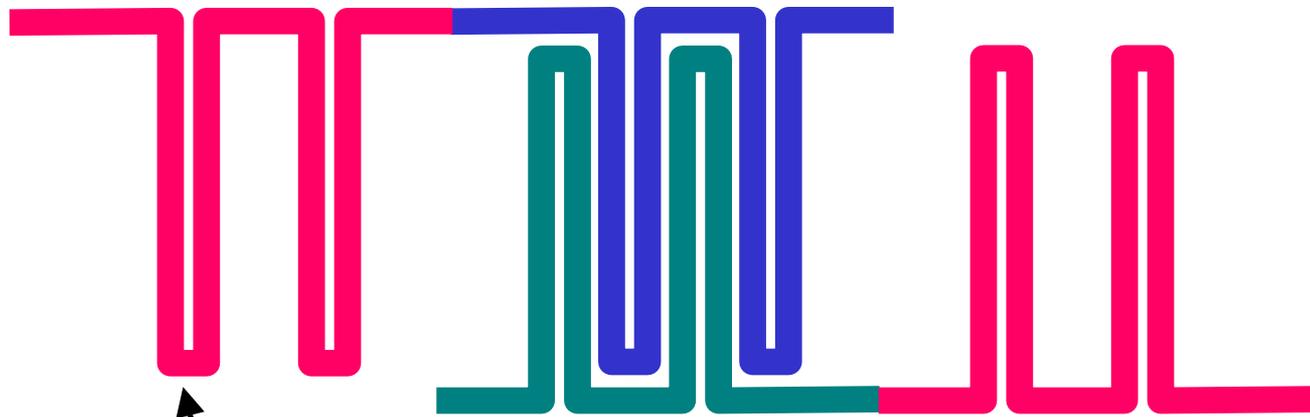
# Layout: Place Sense Resistors



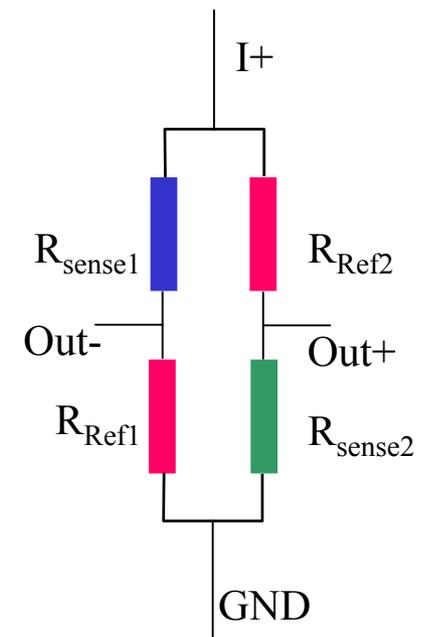
The two sense resistors are interwoven to allow them to sense the same region and experience the same excitation field.



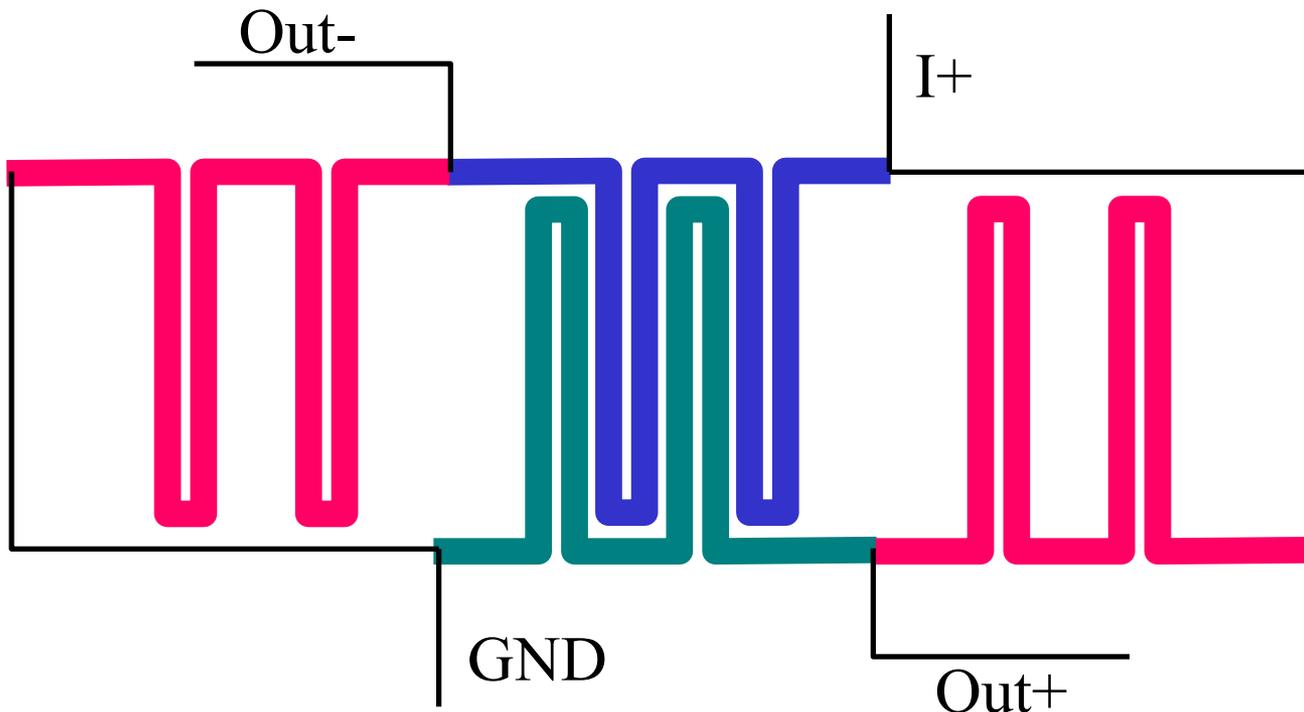
# Layout: Place Reference Resistors



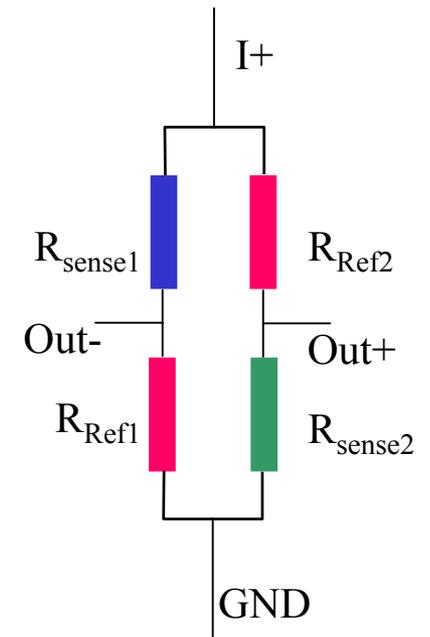
Add two identical  $R_{ref}$



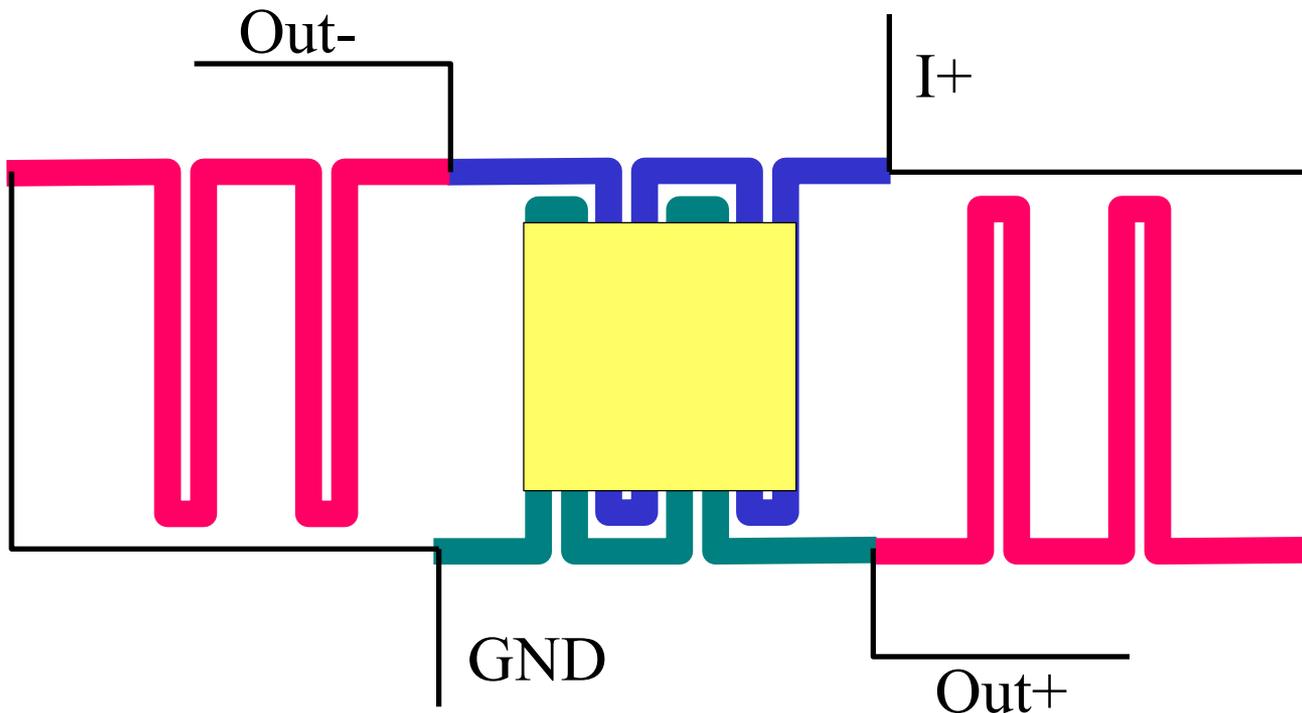
# Layout: Add AI Interconnect



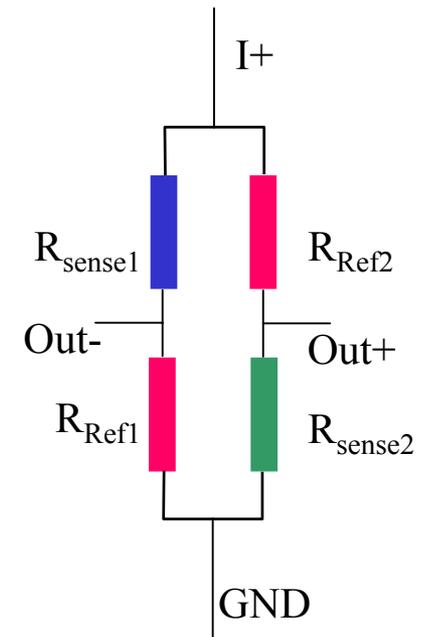
Add interconnection metal (Aluminum)



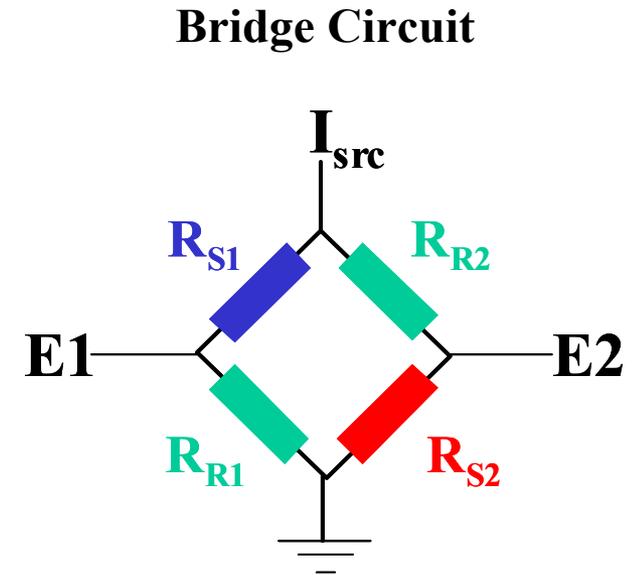
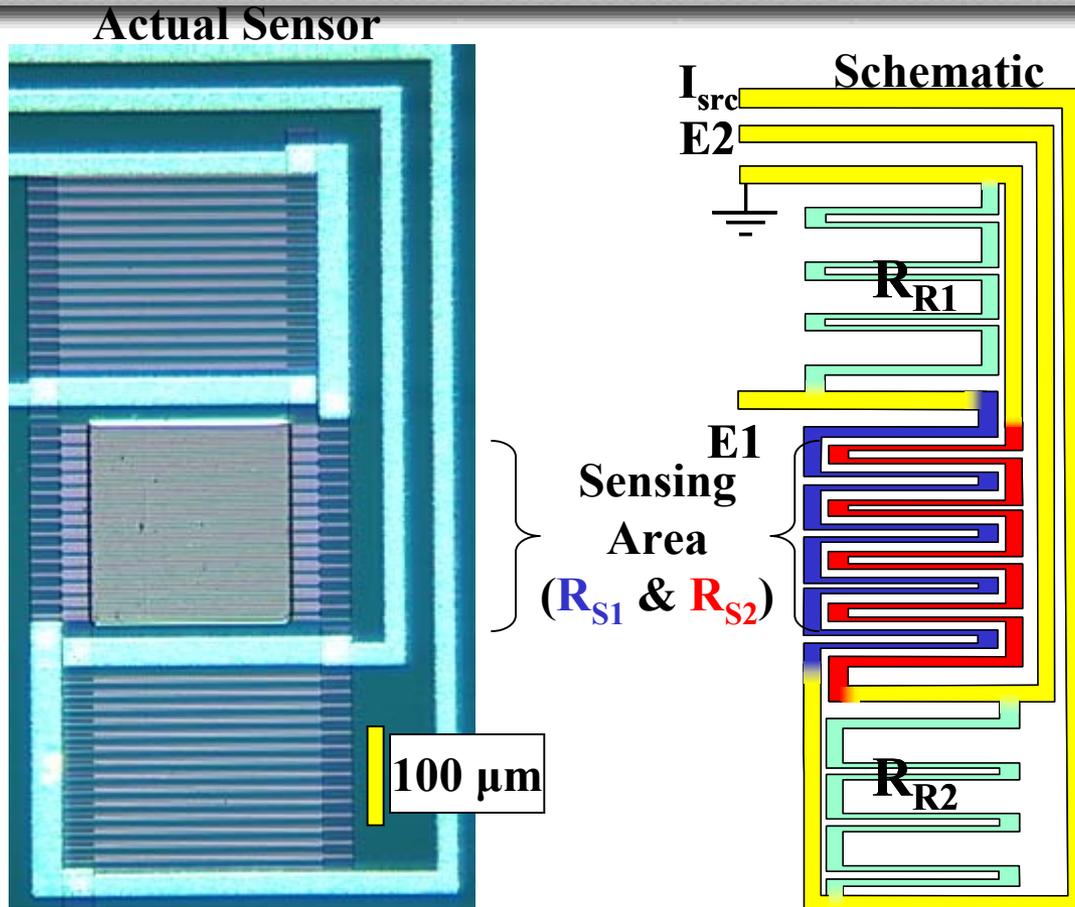
# Layout: Add sense region



Define sensing region by selective etching and surface chemistry



# GMR Chip Design



- A bridge configuration is used for the GMR circuit to reduce baseline drift. The two sensor bridge doubles the signal of the traditional single sensor bridge.

- Sensing GMRs – interdigitated  $R_{S1}$  and  $R_{S2}$
- Reference GMRs –  $R_{R1}$  and  $R_{R2}$

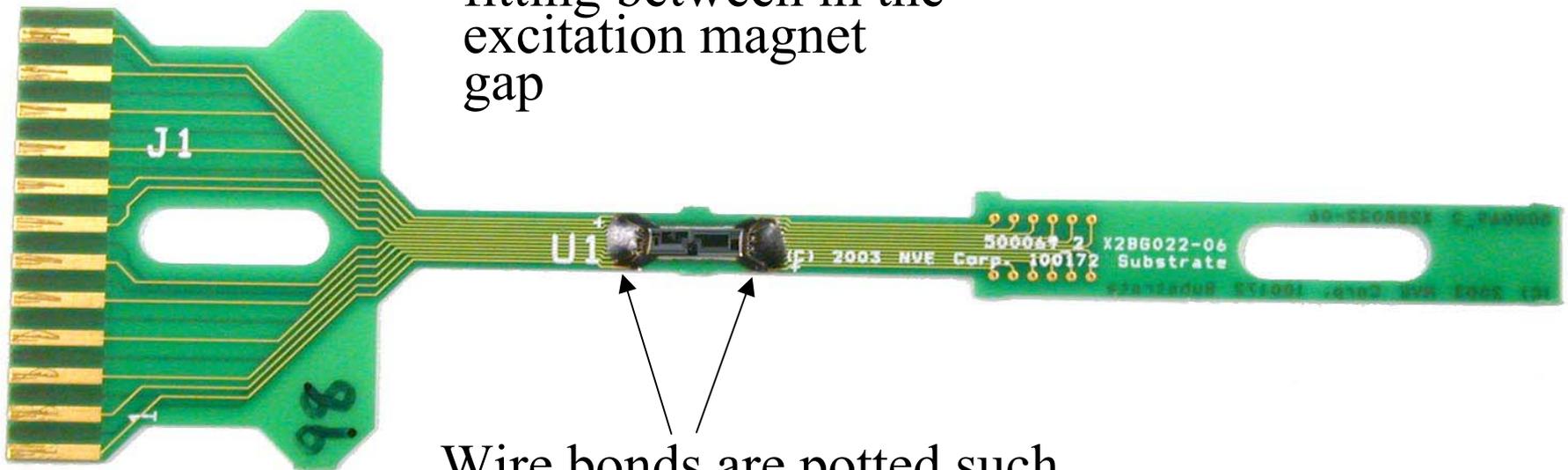
# Die-Holding Printed Circuit Board

## “Diving Board”

### Design Basics

24 pin surface mount  
edge connector

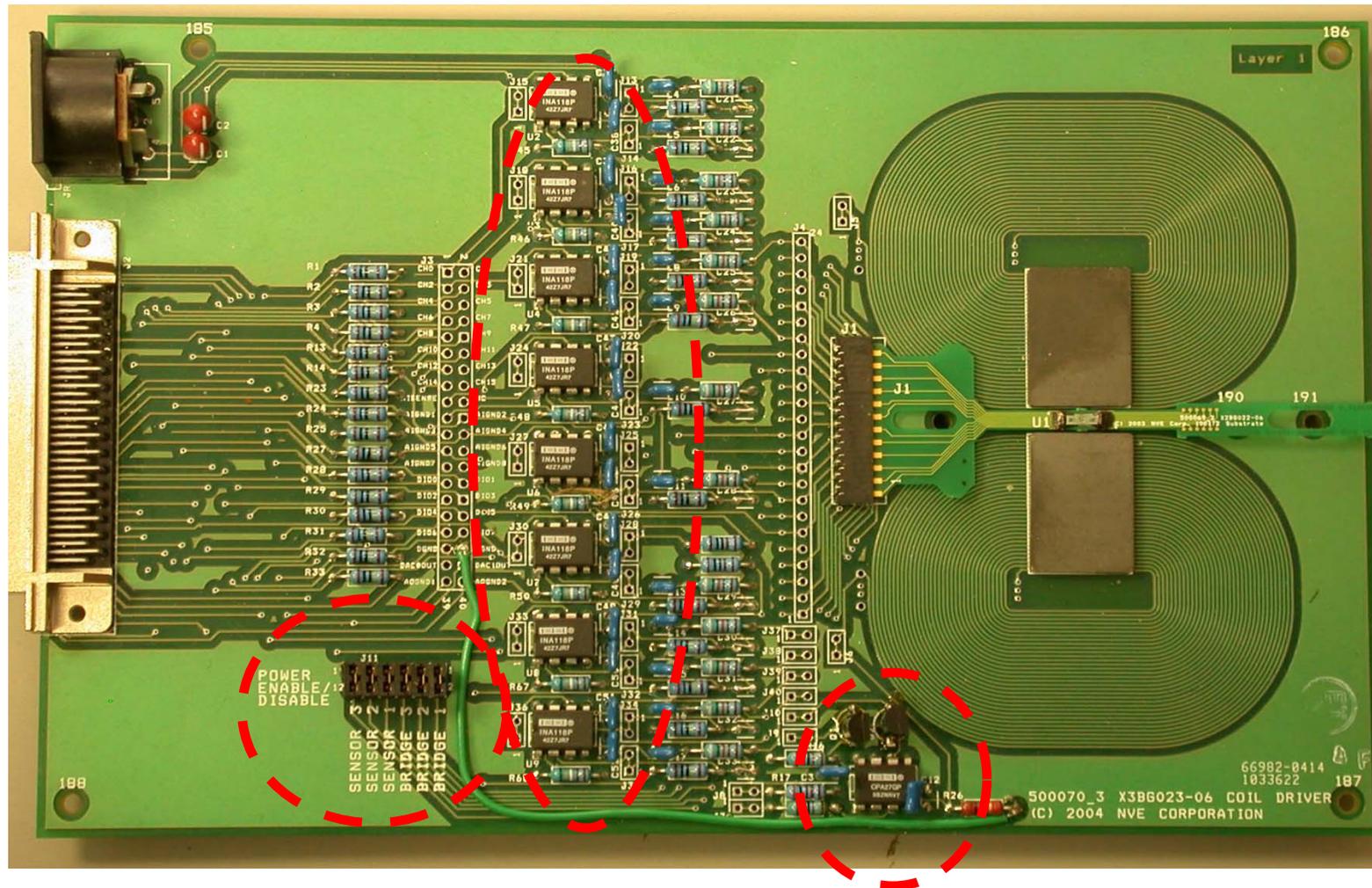
Narrow die area for  
fitting between in the  
excitation magnet  
gap



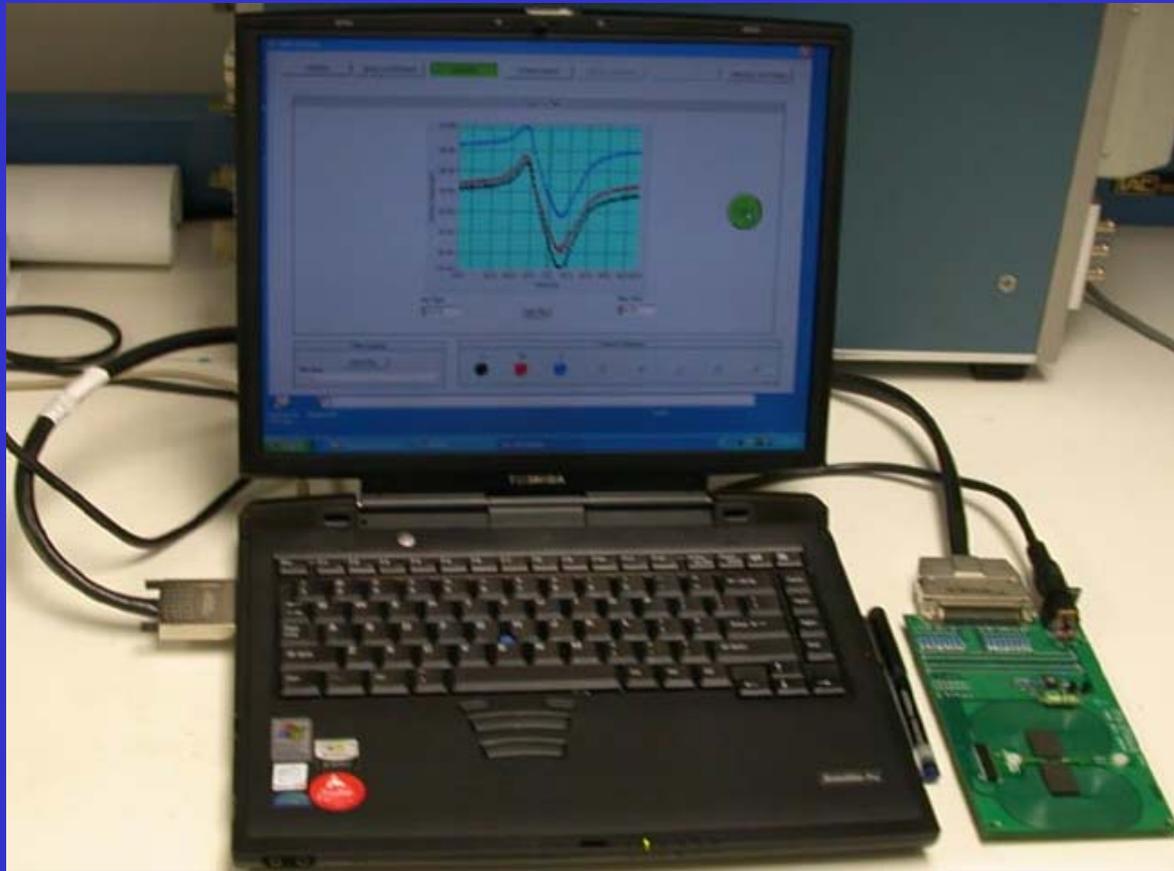
Wire bonds are potted such  
that sense pad is still exposed

# Magnetic Excitation Module

- 8 On-board signal preamps
- Jumpers for sensor channels
- Jumpers for coil driver



# BioMagnetIC System



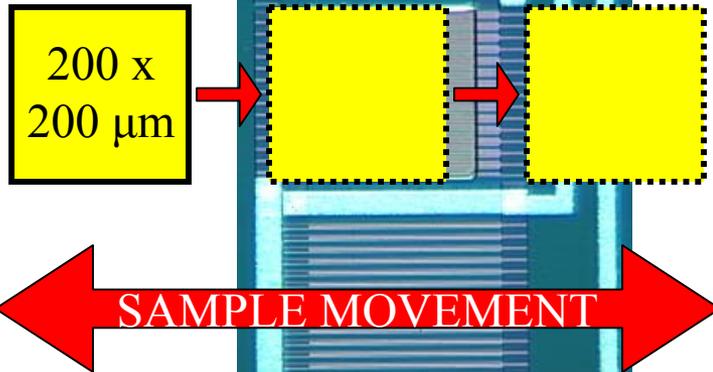
- A to D card in laptop
- Pocket-sized excitation module
- Disposable sensor cartridges
- Adaptable device development platform
- R vs. H plots
- Vout vs. time

# Magnetic Bioassay: Scanned Sample Mode

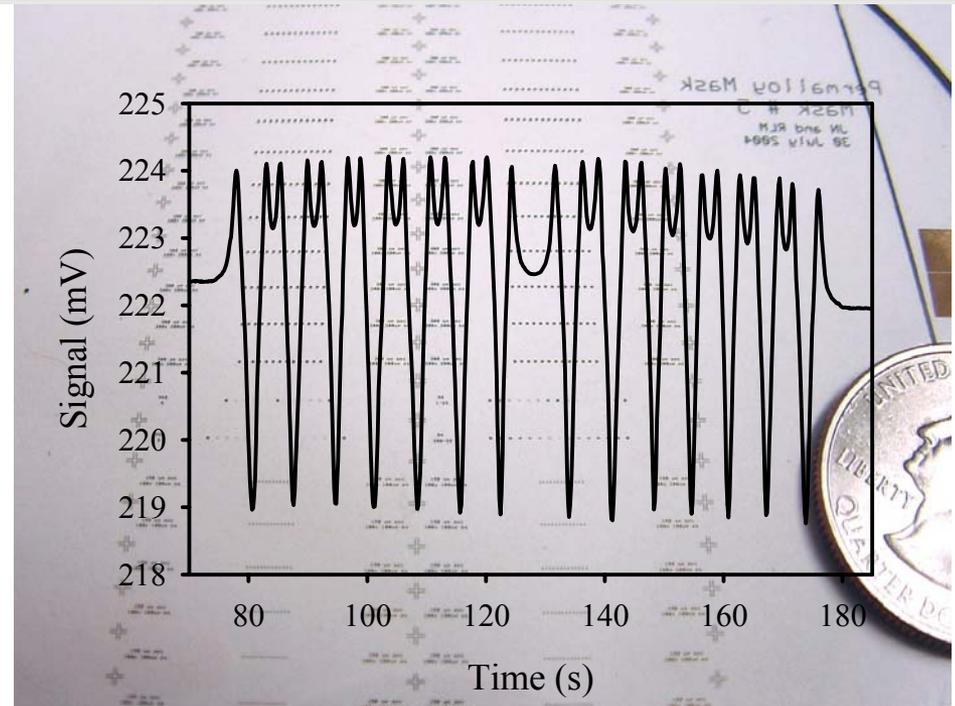
# Sample Stick Experiment

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Move sample across GMR sensing area

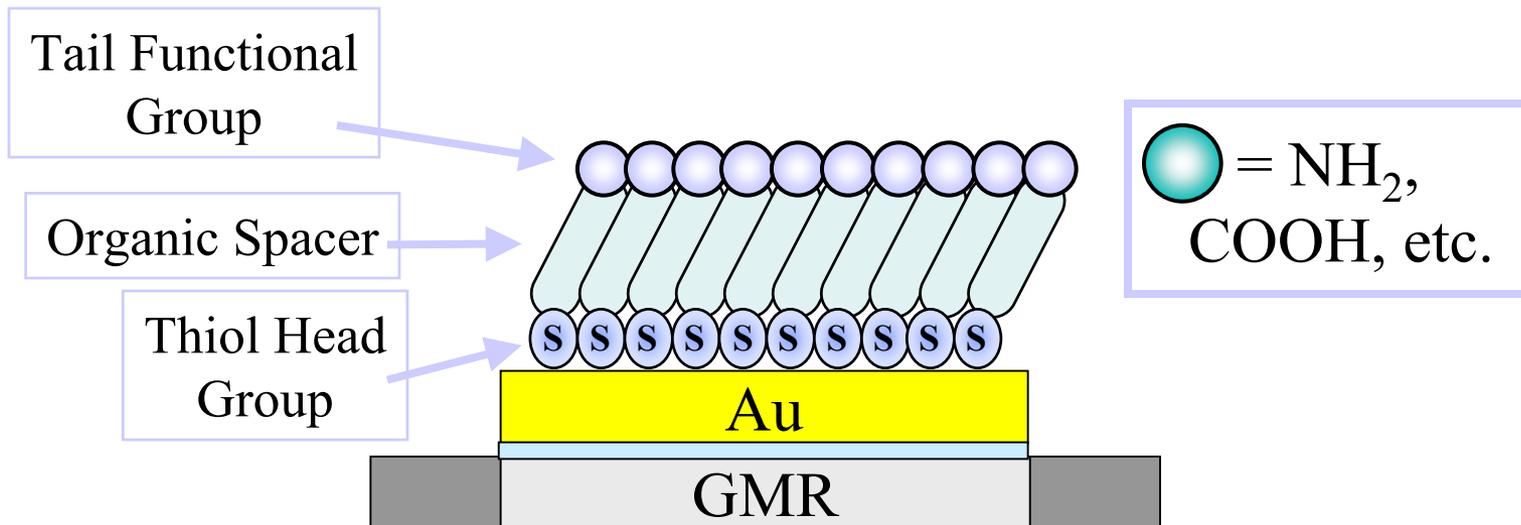


- External field at 150 Oe
- Sample is held near the GMR ( $\sim 50 \mu\text{m}$ ) and moved across sensing area



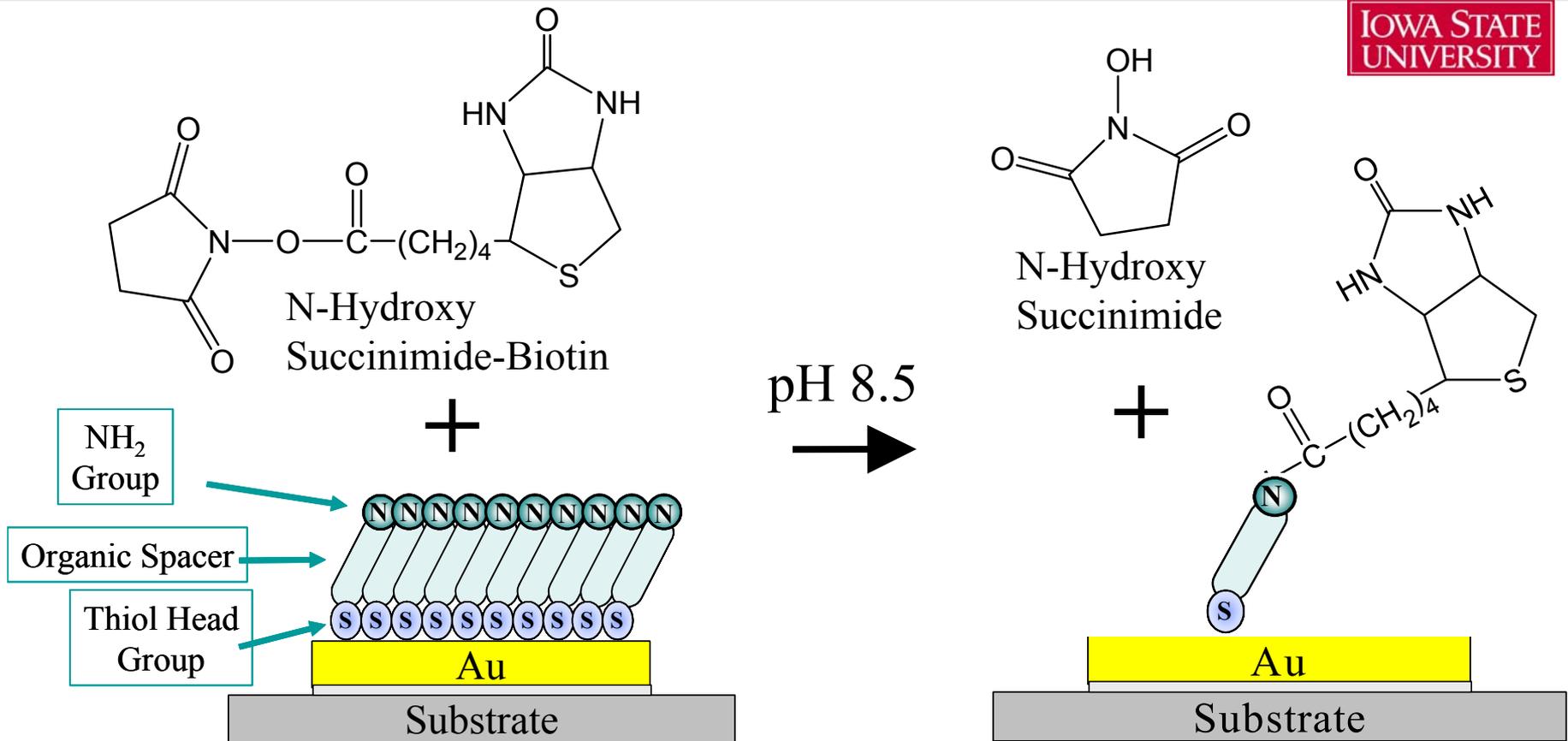
# Surface Functionality & Versatility

- Chemistry of self-assembled thiolate monolayers on gold is extremely versatile
- The thiol head group binds strongly to the Au surface
- The tail functional group can be tailored for desired sensor specificity



# Surface Biotinylation

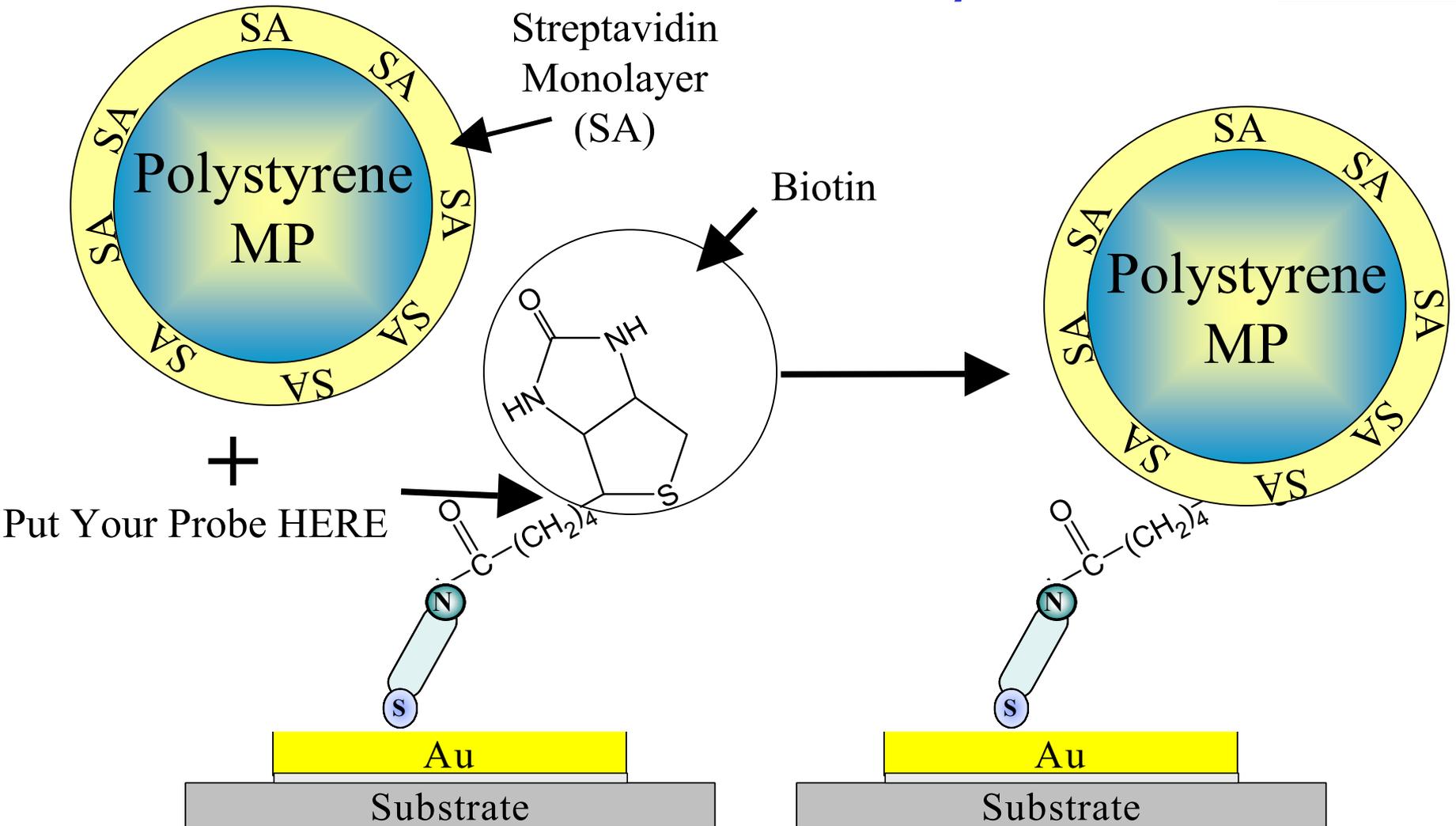
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NHS-Biotin reacts with primary amines to form an amide bond.

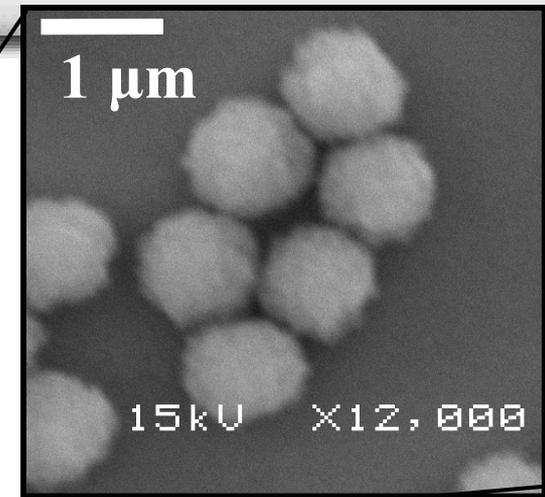
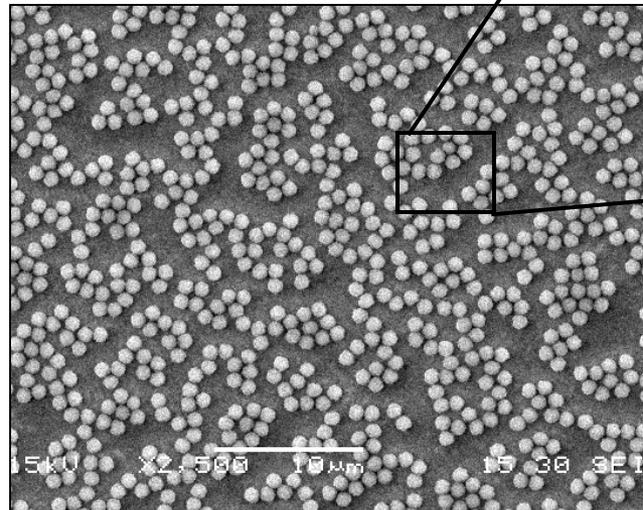
# Assay Labeling with Magnetic Particles:

## Biochemistry



# Magnetic Particle Binding Optimization

- Dynabeads®  
MyOne™ Streptavidin
- Superparamagnetic polystyrene beads (26% iron)
- Monolayer of streptavidin covalently attached to bead surface
- 10 mg/mL in phosphate buffered saline (pH 7.4, 0.01% Tween 20, 0.09% NaN<sub>3</sub>)

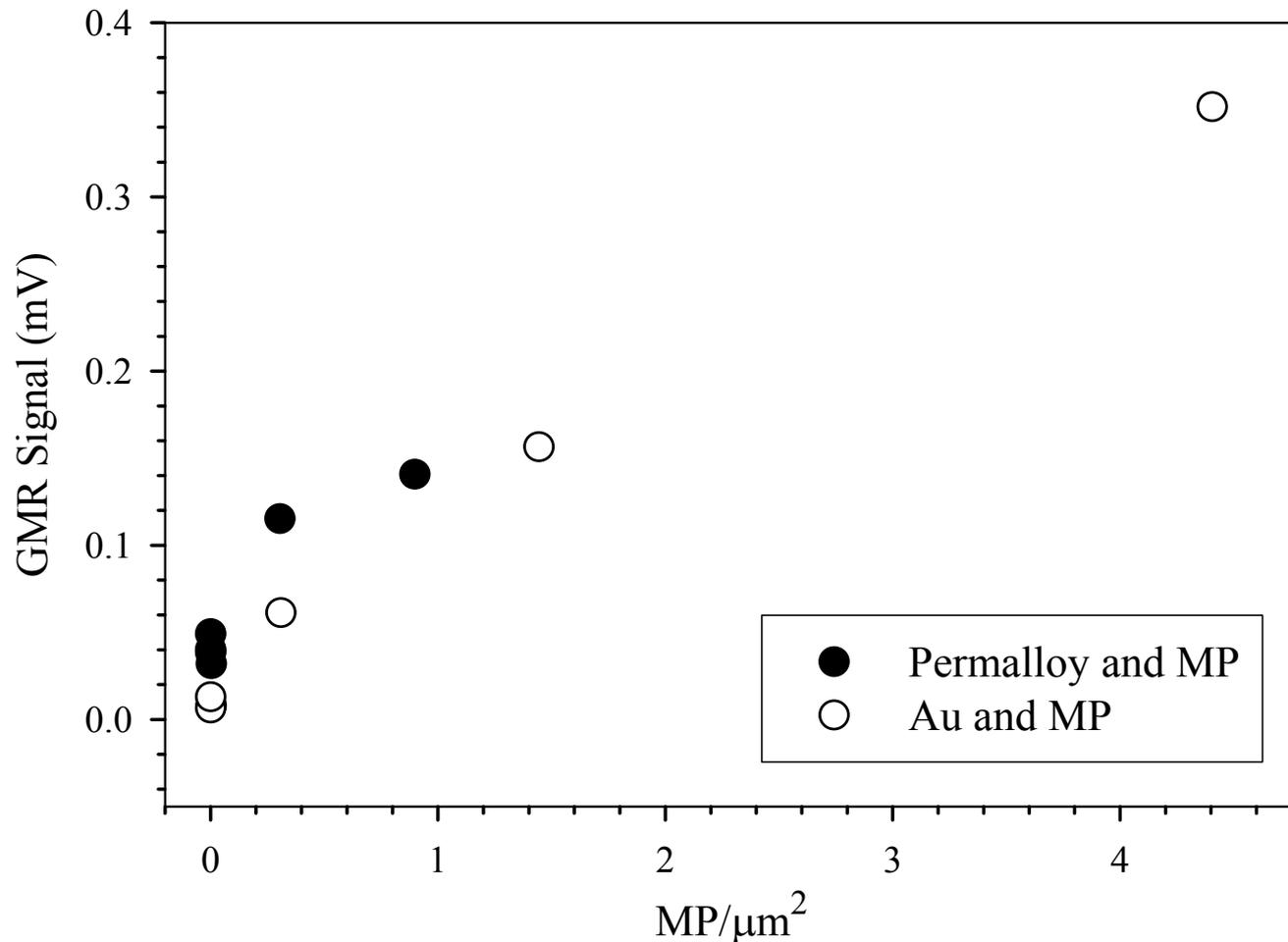


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**MP binding was studied to optimize binding procedures and conditions**

**\*20 μL of 2 mg/mL MP on each sample**

# GMR Signal vs. MP Concentration



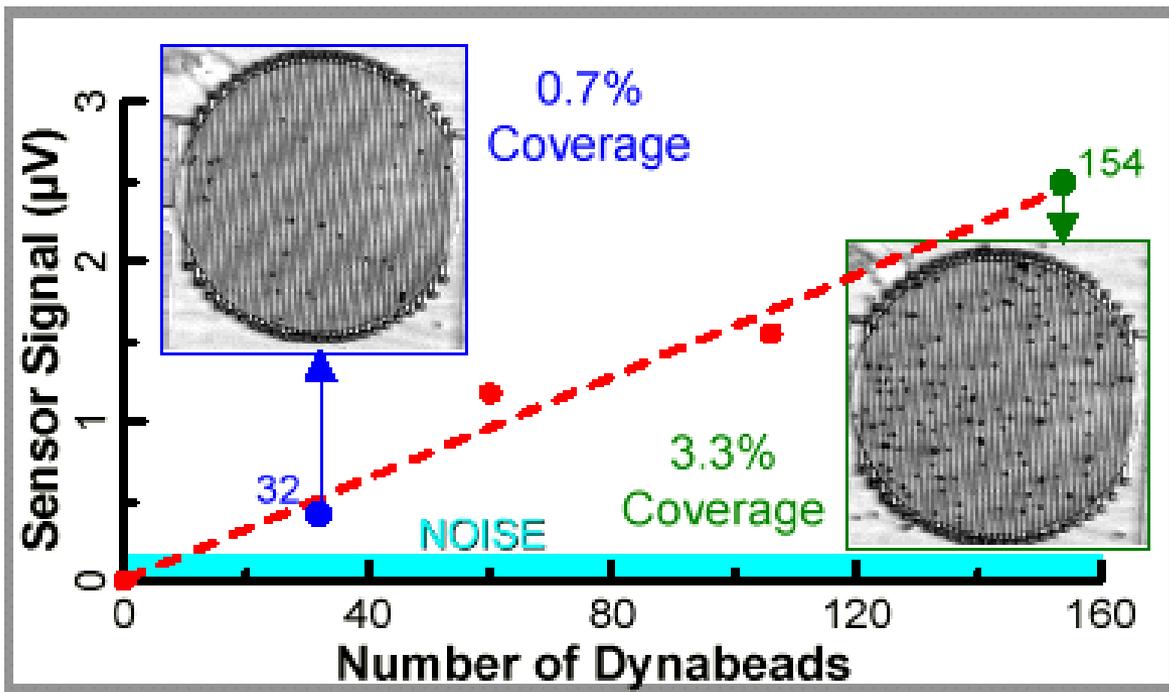
- **Limit of detection (3x SNB) is 0.005 MP/μm², or 215 MP per gold square.**

Best Sensitivity: Bind directly on detector

# Magnetic Biosensor $\mu$ Array: High Sensitivity

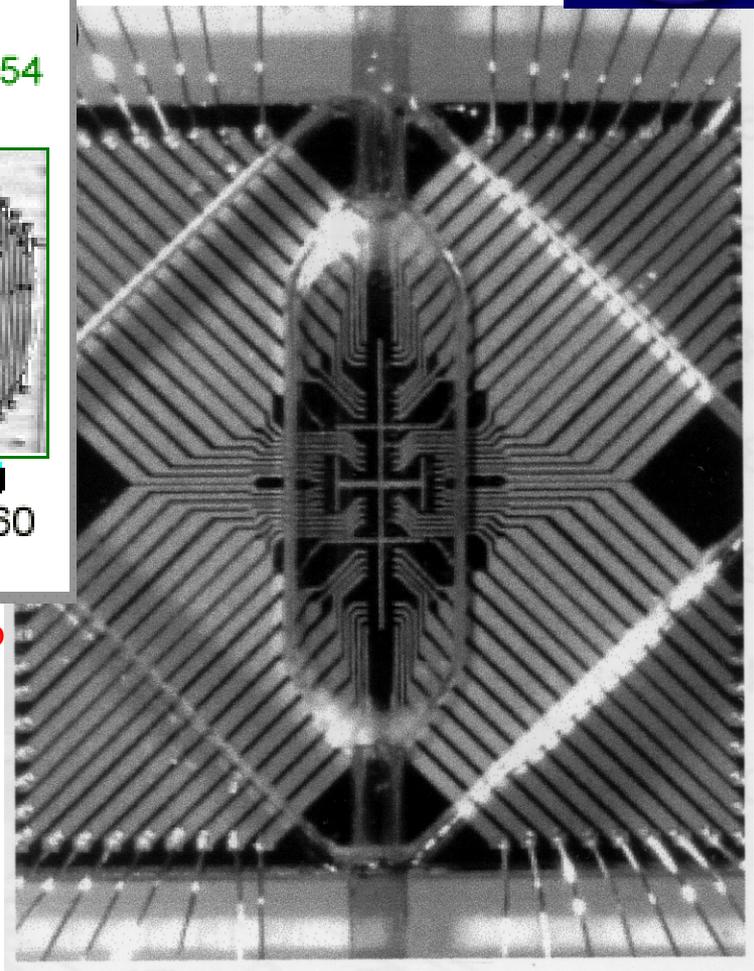


2.8 micron DynalBeads



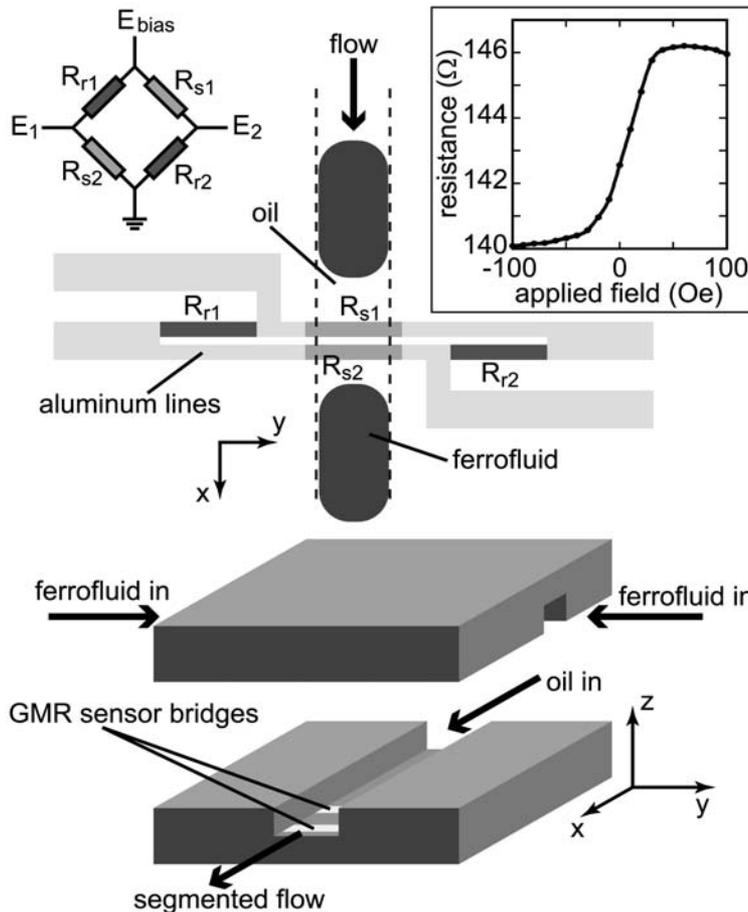
Courtesy L. Whitman, Naval Research Lab

- 1) Single label detection is possible
- 2) >3 decades of dynamic range
- 3) Better than 1 fMolar with fluidics
- 4) Magnetism enhances specificity



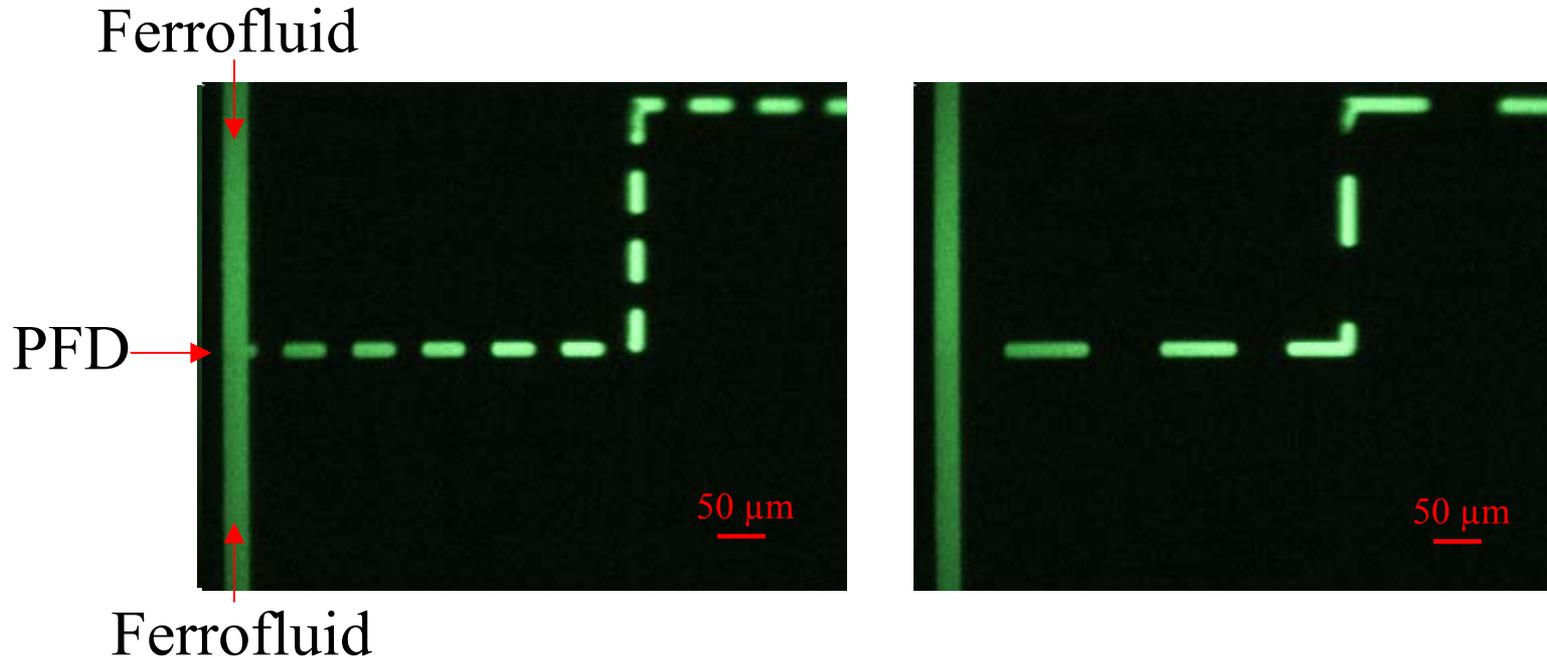
# In-Flow Integrated Detection of Magnetic Droplets

# GMR Sensing of Magnetic Picodroplets



- Picoliter-sized droplets of ferrofluid formed at a fluidic junction
- GMR sensitivity 0.07%/Oe
- Wheatstone bridge configuration

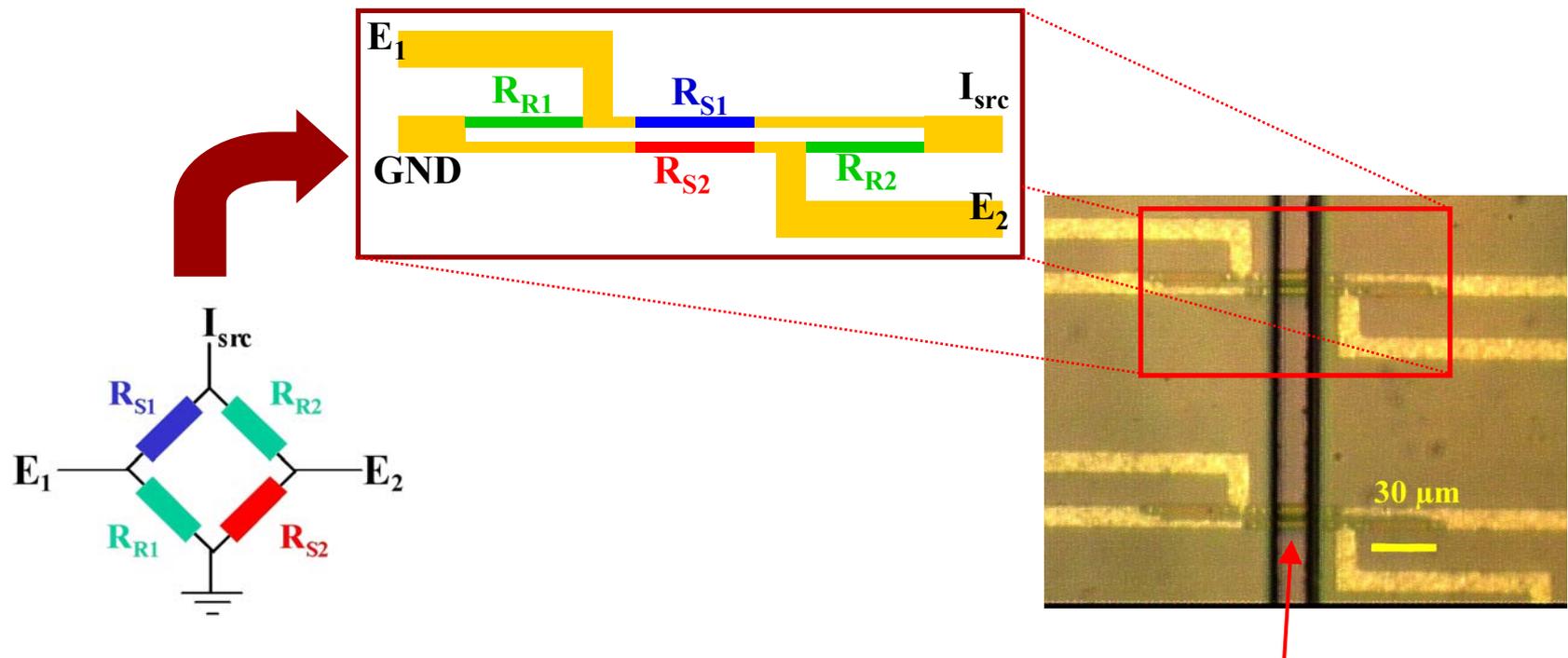
# Ferrofluid Plug Formation



- Flow rate 1.0  $\mu\text{L}/\text{min}$
- Plugs formed at approx. 500 Hz
- Flow rate 0.2  $\mu\text{L}/\text{min}$
- Plugs formed at approx. 50 Hz

# GMR Sensor Architecture

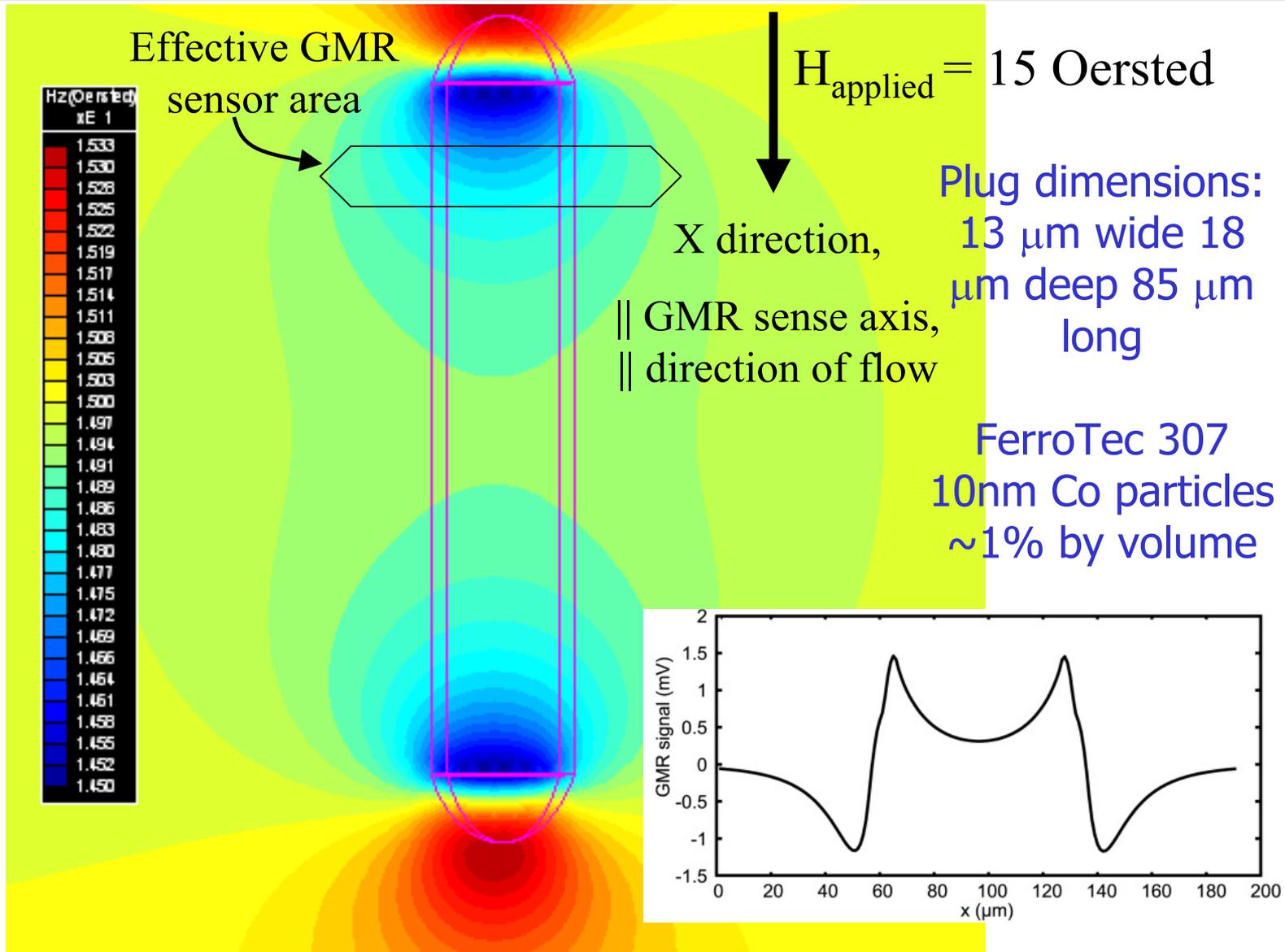
- Two reference and two sensing GMRs configured as a Wheatstone bridge



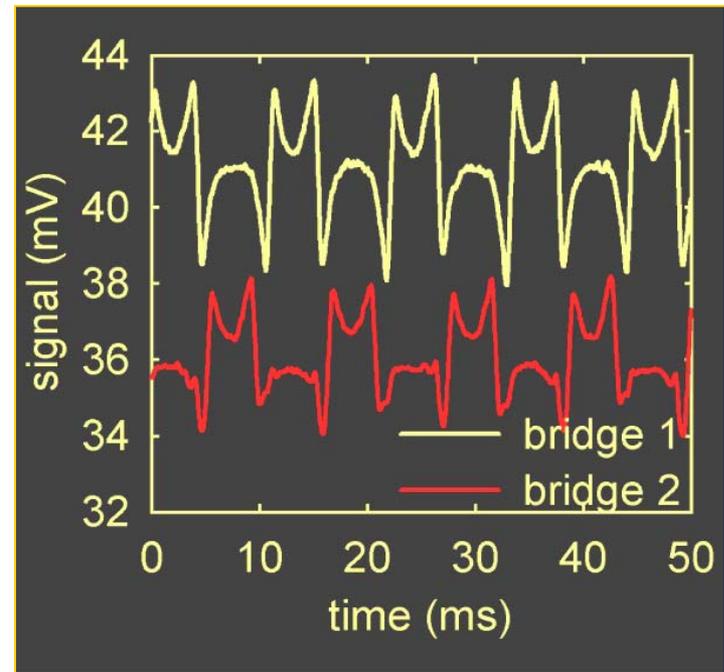
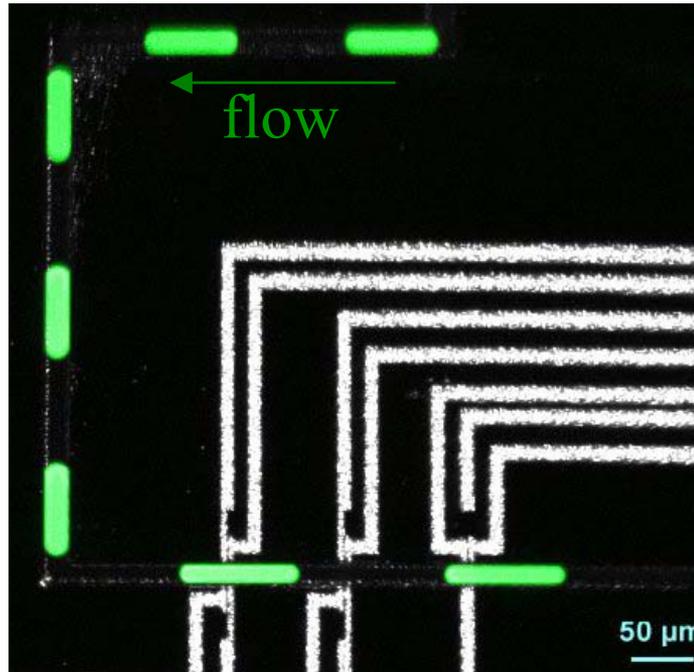
- Channel passes over sensing GMRs

# $H_x$ in Sensor Plane, Flowing Ferrofluid Plug

Simulation using "Amperes" magnetic modeling software



# Direct Flow Velocity Monitoring

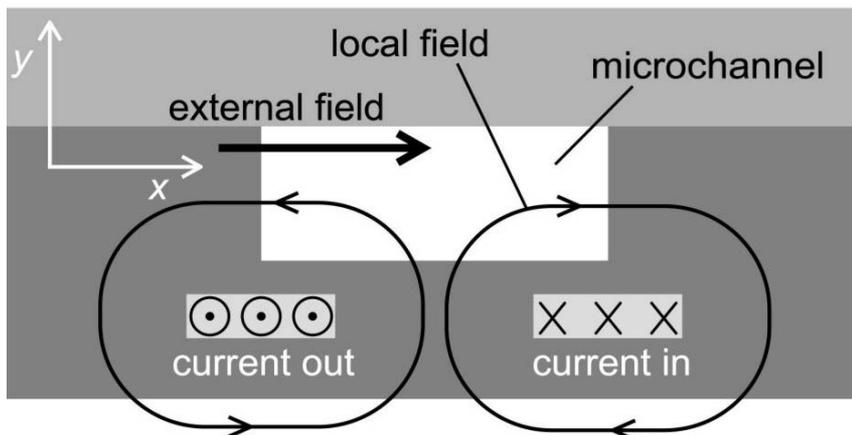
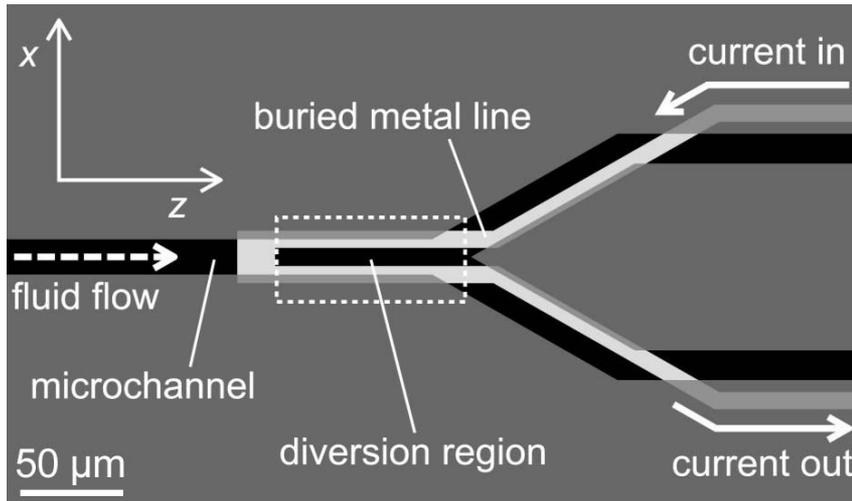


Excitation field 15 Oe; Flow rate 250 nL/min; 1.2% magnetite v/v

Velocity determined by cross-correlating the signals from two bridges

# In-Flow Manipulation of Magnetic Particles

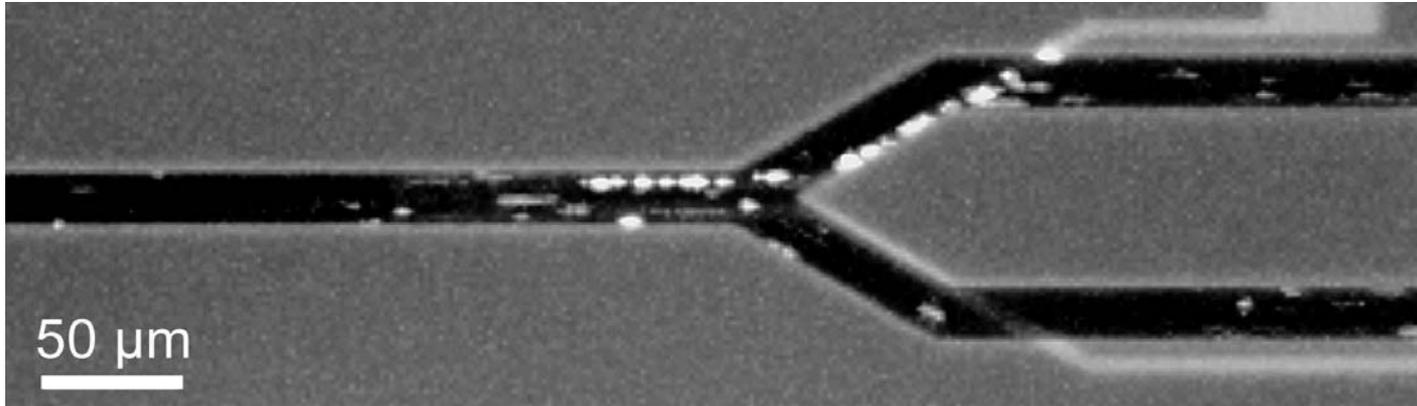
# Diverter Design and Fabrication



- A uniform external field magnetizes particles
- Current lines induce field gradients of  $10^2$ - $10^3$  T/m
- Resulting force diverts particles to a desired channel

# Magnetic Flow Sorting Experiments

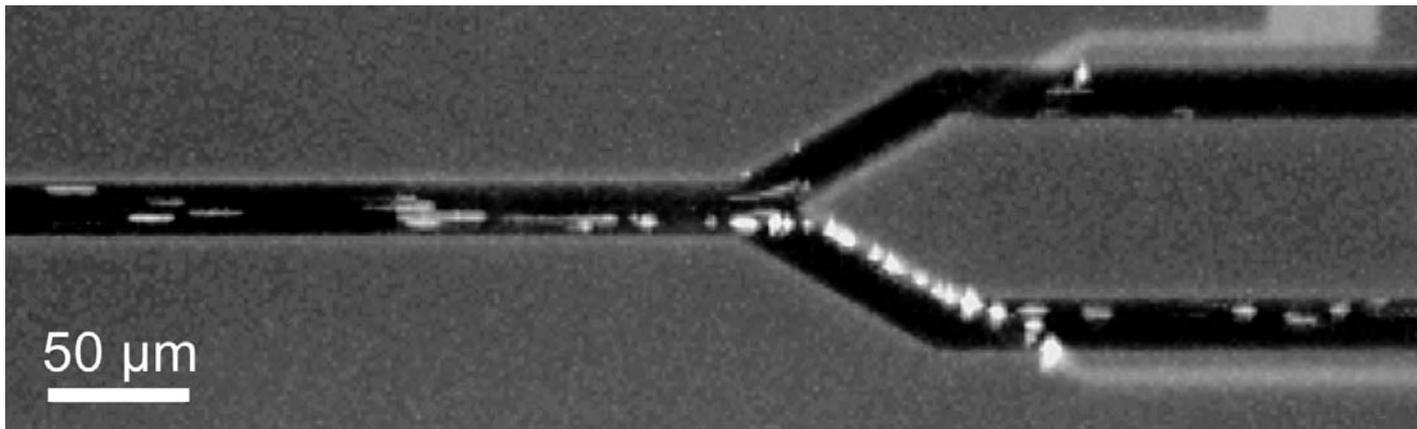
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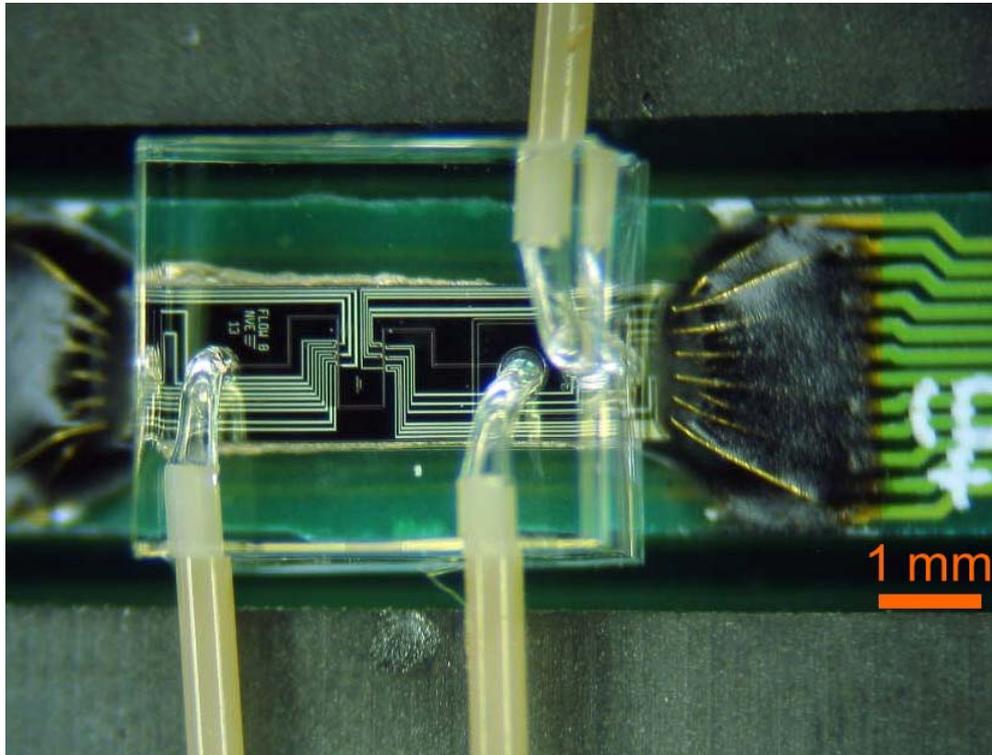
Bangs Labs, 28%  
magnetite, 1  
 $\mu\text{m}$

Flow rate: 6  
nL/min

85% of the beads  
in desired  
channel

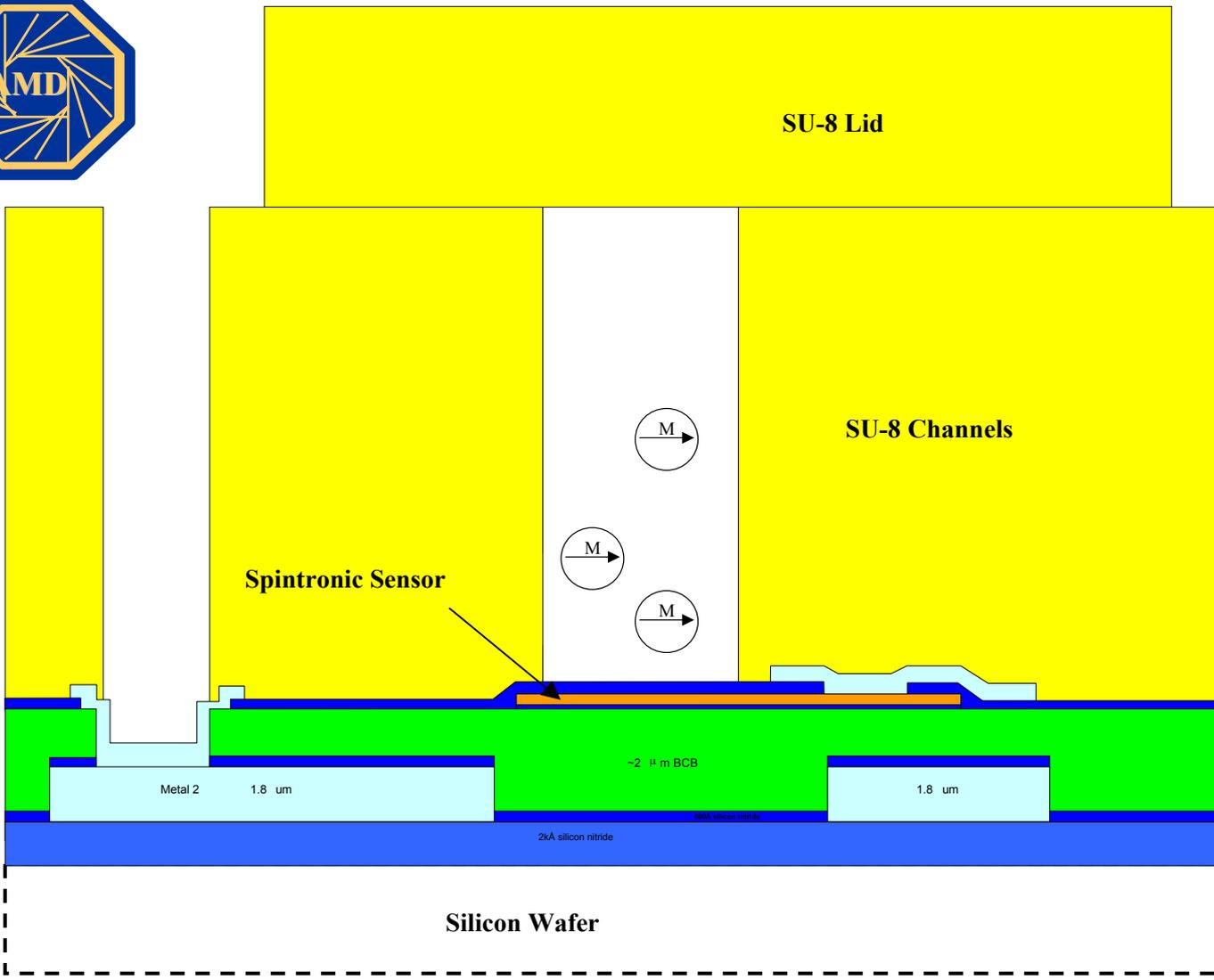


# Device Packaging with PDMS



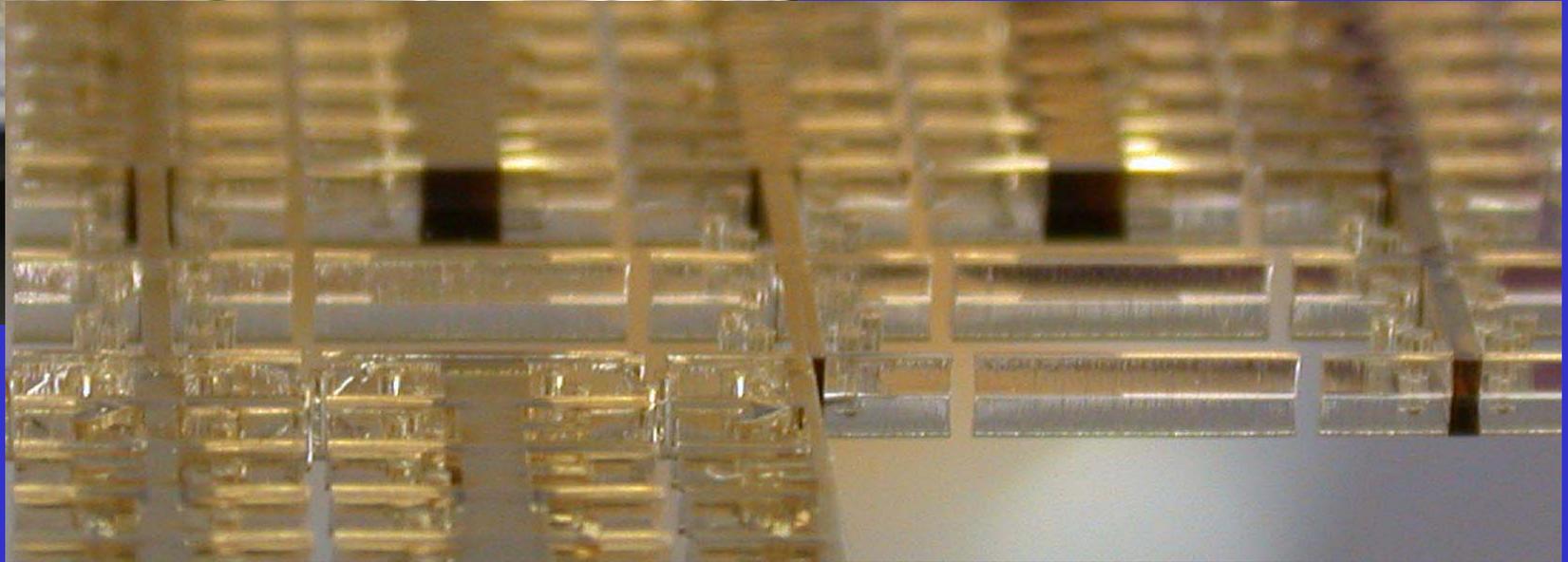
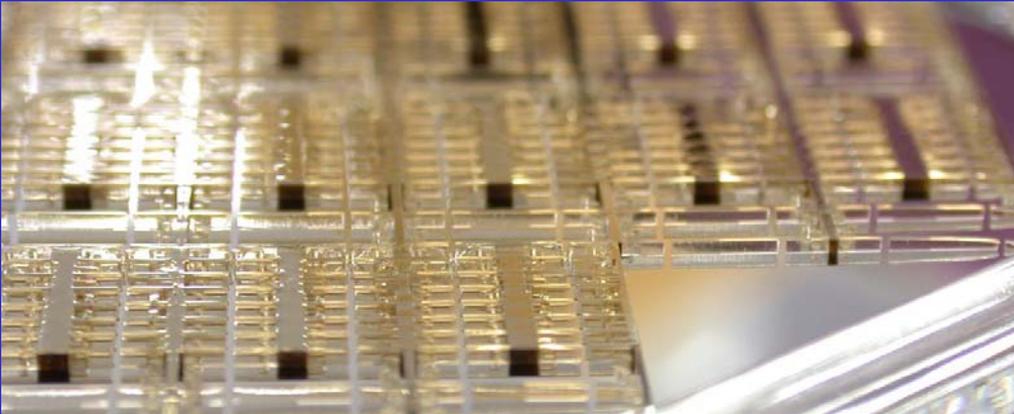
- Electrical connections
- PDMS Fluidic connections
- Optical access

# SU-8 Lid using wafer-scale bonding method



# SU-8 on GMR wafers

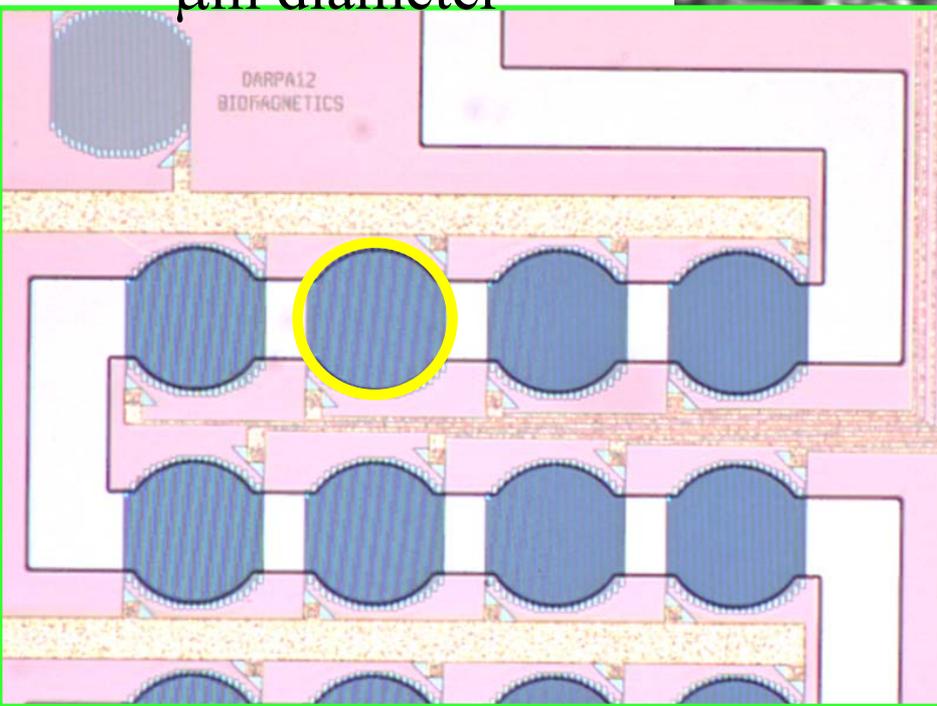
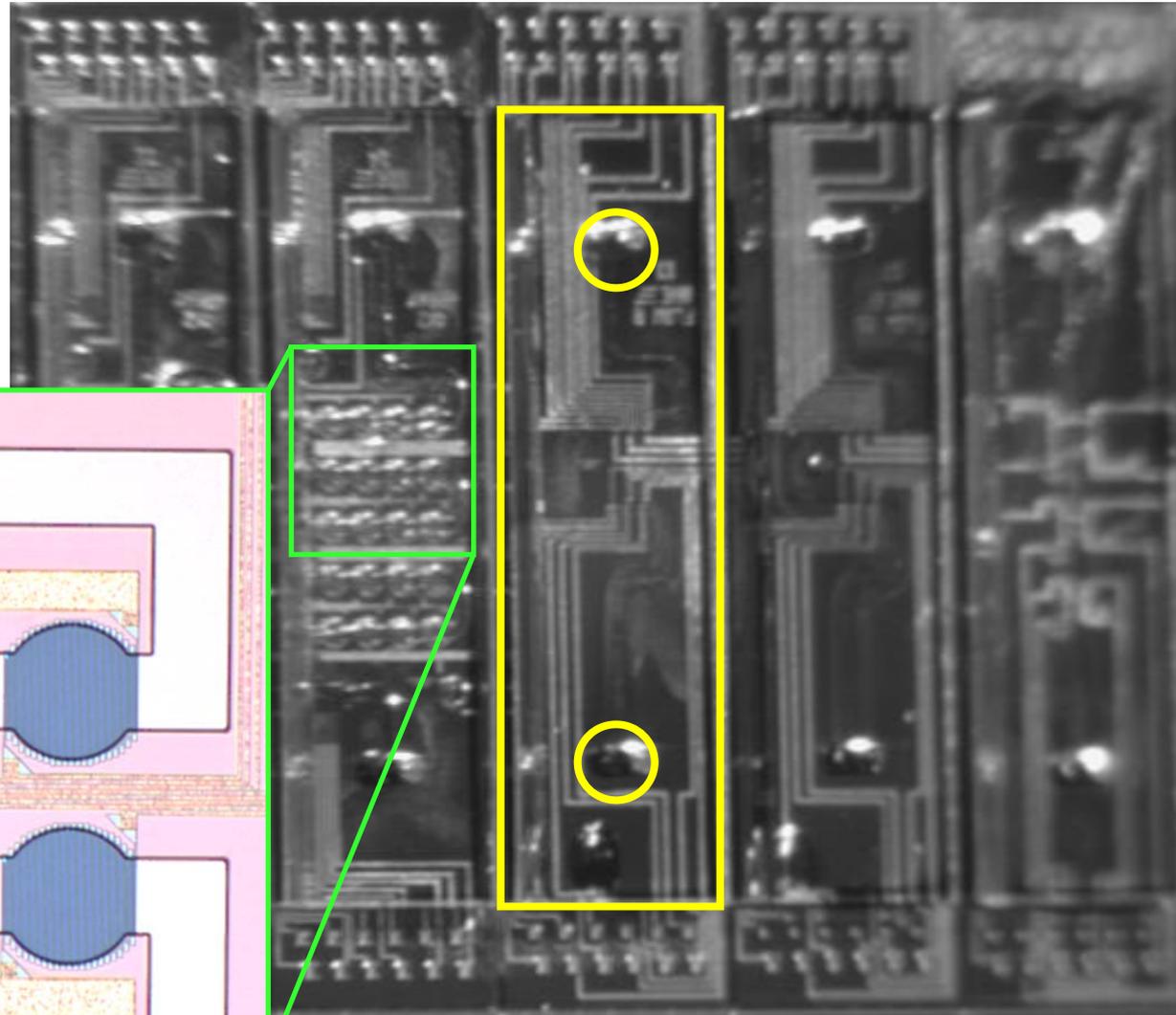
Up to 860  $\mu\text{m}$  thick  
Ultra-high aspect ratio  
Wafer level processing  
Rapid prototyping  
Adding: Multi-level, pick & place



# Covers with 100 micron ports on the wafer



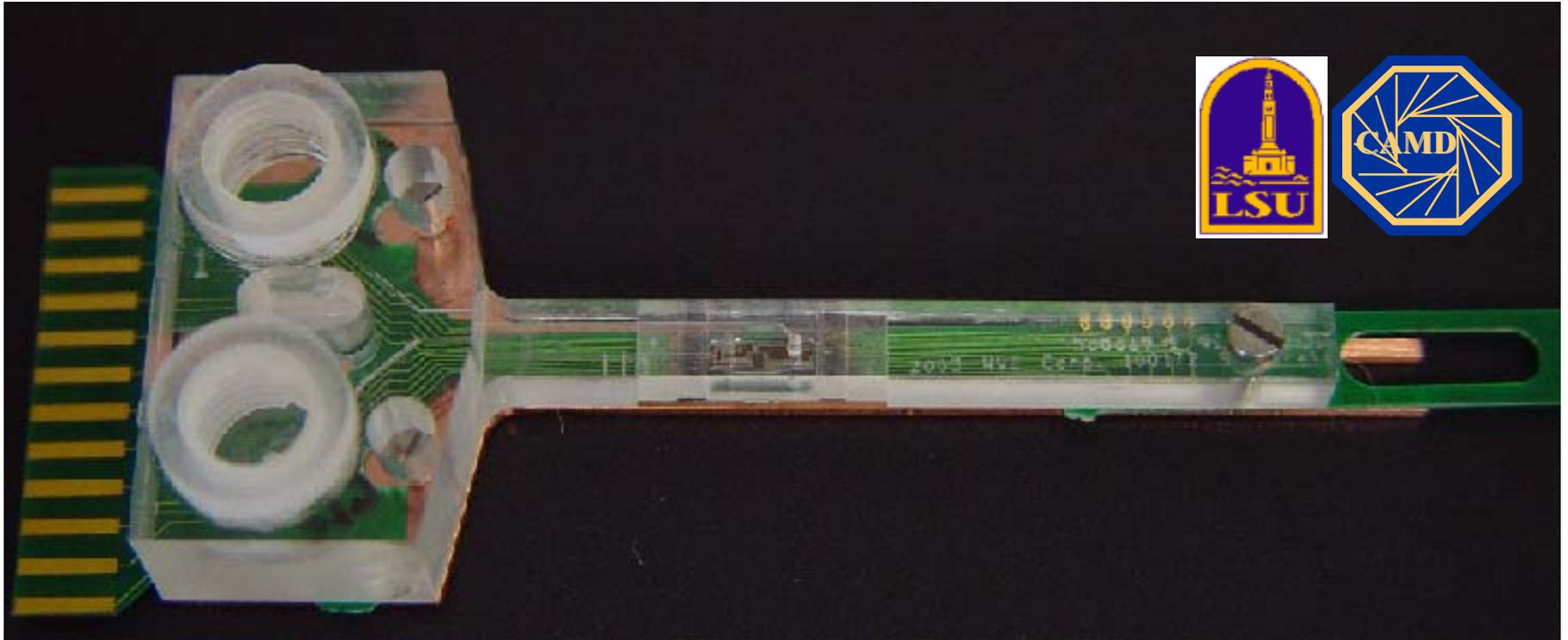
20-sensor array, 200  $\mu\text{m}$  diameter



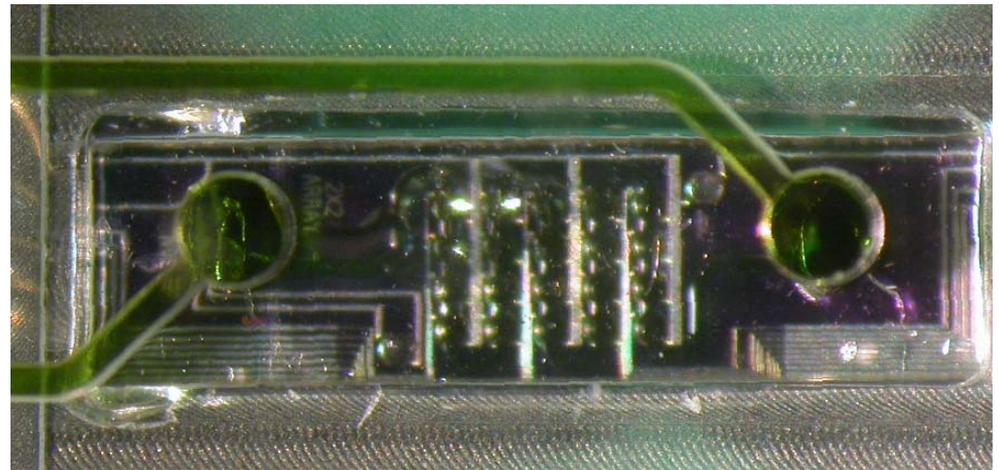
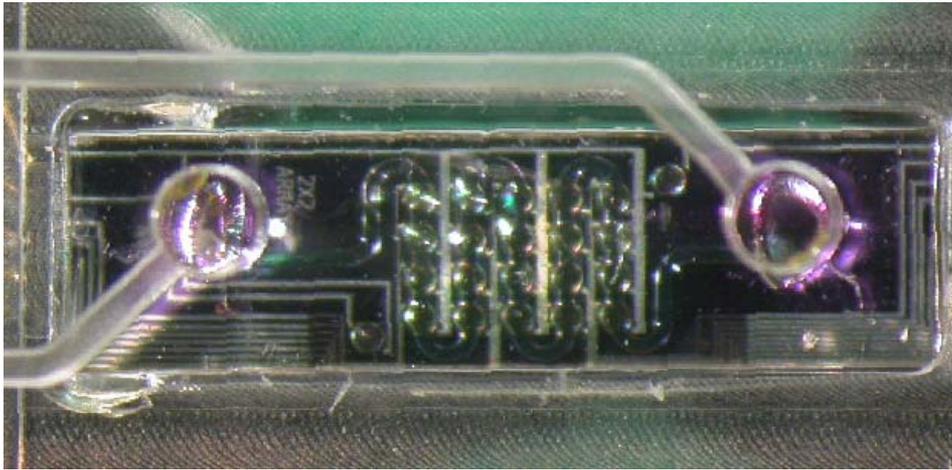
# Dice, mount on circuit board, wire bond



# Ready to use fluidic / GMR chip



# Ready for flow: before and after pumping liquid



# Detector Comparison

Magnetic	Photonic
Little background	Many light sources
Array is embedded in chip	Array must be scanned mechanically
Integrated single label detection is natural	Single label detection requires sophisticated optics
Less "colorful"	Narrow-band labels
Fields are not easily steered	Photons can be directed, diffracted, etc
Challenging fluidics integration	Challenging fluidics integration

**Both are more sensitive than electrochemical**

# Comparison with Electric Manipulation

Magnetic	Electrophoresis or E-field
Not limited by buffers	Need special buffers
Little spurious magnetism	Many charged items
External magnetic field source	Easiest device to fabricate
Challenging fluidics integration	Challenging fluidics integration

# Near-term Technical Goals



- Demonstrate Single Particle Detection
- Demonstrate a flow cytometer using magnetic labels and sorting

# Next Generation

## Ultimate:

### **Molecular counting and bead-by-bead manipulation**

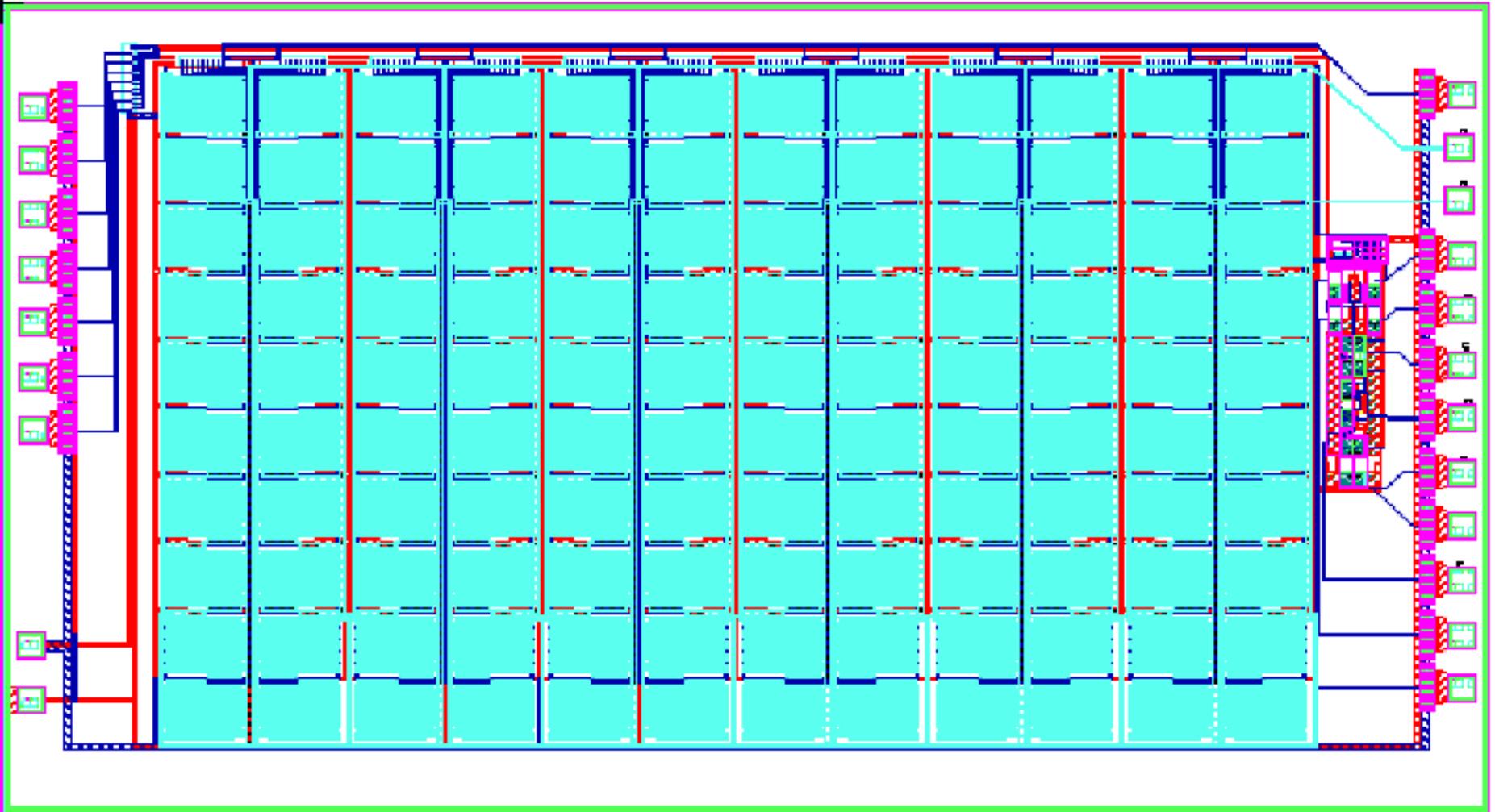
- Reduce label size to ~10 nm (more difficult detection)
- Use magnetics to enhance assay dynamics (mixing, capturing, binding hybridization, sorting)
- Increase system complexity by combining components
- Increase assay complexity with multiple analytes, sensors, and channels.

# Biosensor array parameters

- First commercial array will be for Veterinary screening, ~ 20 sensors, 200 micron diameter, immunoassay (Seahawk Biosystems) (disposable cartridge with sensor inside)
- Largest array without integrated circuits is 68 sensors
- High density DNA chip format possible (1,000,000 sensors)
- Currently building a 96 sensor chip with integrated circuit switching

# Integrated Circuit Design: 96-site array

0.35 micron TSMC wafers now in-house



# Disposable sensor vs. sensor instrument

- Permanent sensor could scan lateral flow assays, glass slides, tapes, etc.
- Detection performance is greatly influence by proximity (microns)
- Cost per sensor die is low,  $\ll \$1$
- Typical GMR product retail cost  $\sim \$1 - \$3$

# Possible Applications

- Portable high performance bioanalytical system for military
  - Food and water safety (detect e-coli, salmonella, ..)
  - Chemicals that can be bound magnetically
  - Disposable DNA chip, gene expression
  - Immunoassay
- 
- Best apps would benefit from high-volume low-cost model

# Summary

- Integrated microfluidic tools for detecting and sorting have been demonstrated
- Detection of immobilized and flowing labels demonstrated with wide dynamic range ( $> 3$  decades)
- May open new applications in single molecule detection
- Manufacturable integrated microfluidics
- Magnetic detection system is ideal for high volume, low cost applications, and where portability and ruggedness are important (motivates the military funding)

[www.nve.com/~markt/biomagnetics](http://www.nve.com/~markt/biomagnetics)

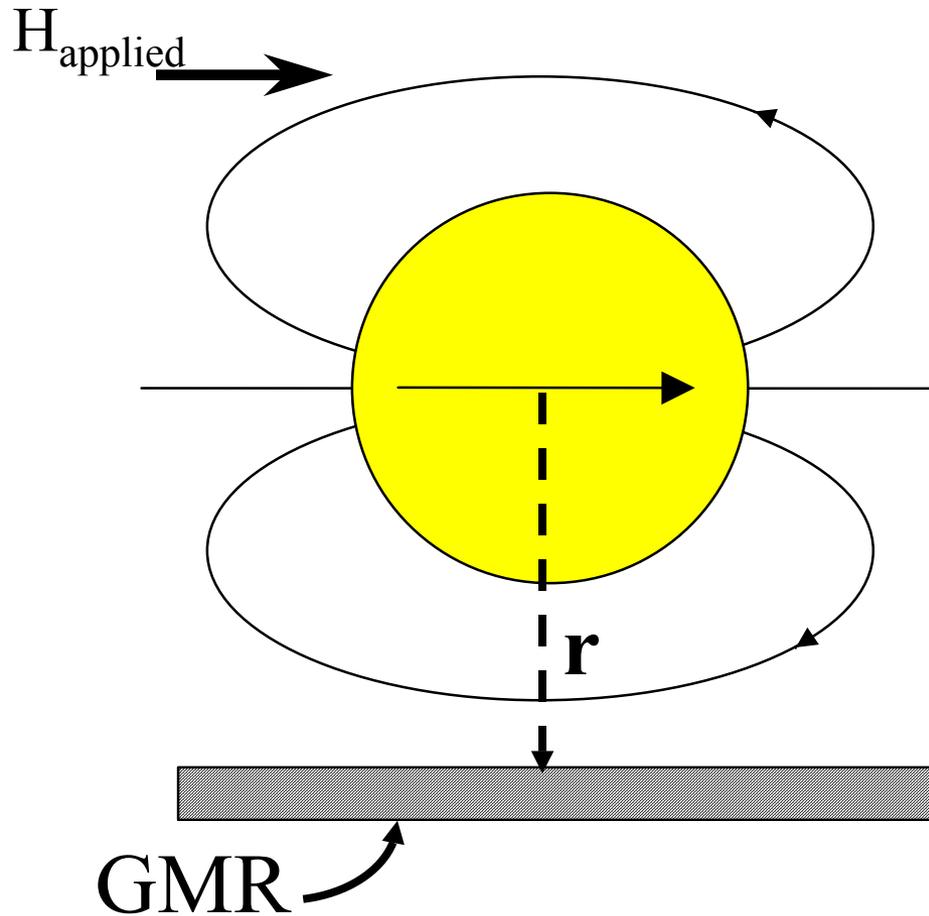
# 10 nm Labels should be detectable

- Signal to noise is fine, hard to ascertain label location
- Dynamic range decreases with label size
- Some assays are better off with much larger detectors
- Get best of Dynamic Range and detection of small labels with an array of detectors
- Technical limits on small end are due to thermal magnetic stability of labels and detectors, fabrication, and characterization challenges, and chemical stability of labels.

# Groups working on nanolabel detection

- S. Wang, Stanford
- M. Johnson, NRL
- P. Freitas, Portugal
- J. Hormez, LSU
- M. Porter, ISU
- M. Megans, Philips

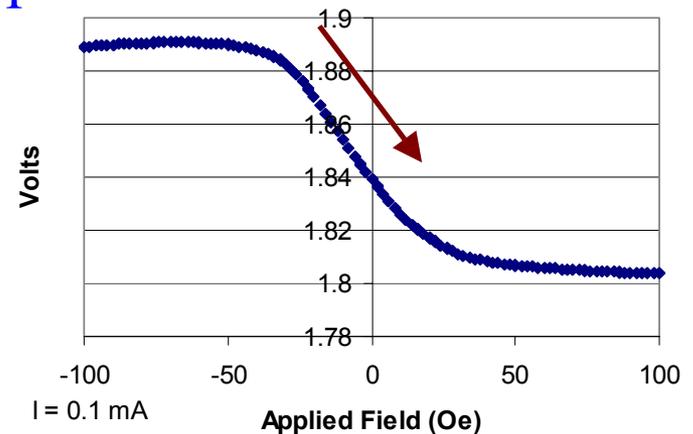
# Stray Fields: in-plane excitation



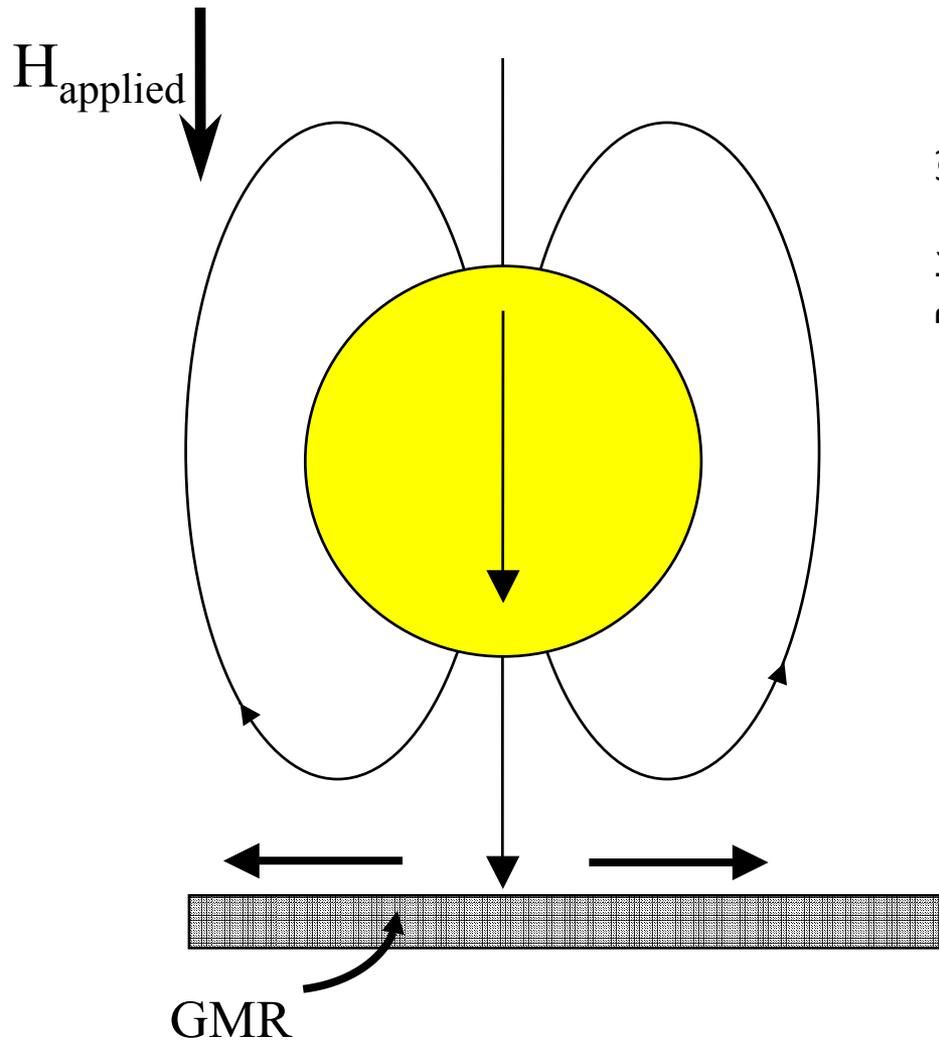
Need a sensor with bipolar output

The detected in-plane stray fields are in one direction

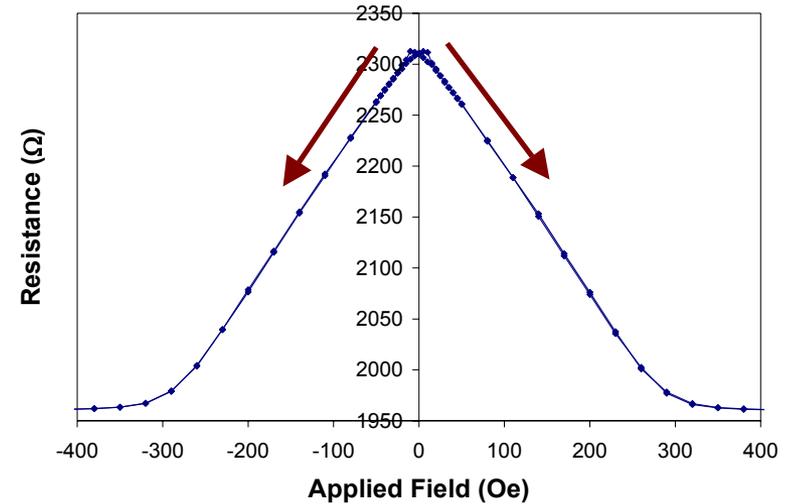
Spin Valve Transfer Function



# Vertical Excitation



## Multilayer Transfer Function



Need a sensor with  
unipolar output

The detected in-plane  
stray fields are in many  
directions